

Identification of a Tool Compound to Study the Mechanisms of Functional Selectivity between D₂ and D₃ Dopamine Receptors

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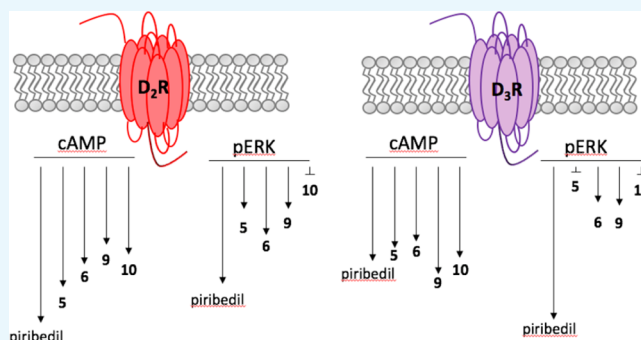
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Supporting Information

ABSTRACT: The search for synthetic selective compounds for G-protein-coupled receptors has provided a myriad of molecules with high selectivity and therapeutic potential. In some cases, however, selectivity is difficult to obtain. For instance, the selectivity ratio is relatively low for compounds acting on D₂ and D₃ dopamine receptors, which are targets of neurodegenerative diseases such as Parkinson's and Huntington's. From a therapeutic point of view, it is of interest the relative recent discovery of biased agonism, which is characterized by different signaling pathways engaged by different compounds acting on a given receptor. The aim of this paper was to investigate whether new piribedil-derived compounds could display higher selectivity for D₂ or D₃ receptor and/or provide biased signaling. The results show that selectivity was not different, but that one of the molecules described here, 5-((4-(pyrimidin-2-yl)piperazin-1-yl)methyl)quinolin-8-ol (**10**), does engage Gi-mediated signaling via D₂ or D₃ receptors, whereas it does not activate the mitogen-activated-protein kinase pathway, which is usually activated by dopamine receptor agonists.



1. INTRODUCTION

Dopamine is one of the main neurotransmitters in the central nervous system (CNS), exerting its functions via five different receptors, D₁ to D₅, that are expressed in different neuronal types and in different regions of the brain. They all belong to the superfamily of G-protein-coupled receptors and are subdivided into D₁-like (D₁ and D₅), which are coupled to G_s and their activation leads to activation of adenylyl cyclase, and D₂-like (D₂, D₃ and D₄), which are coupled to G_i. Apart from mediating changes in one of the main intracellular second messengers, adenosine 3':5'-cyclic phosphate (cAMP), they also engage the mitogen-activated-protein kinase (MAPK) pathway. Dopamine receptors are targets of a variety of neurological and neuropsychiatric diseases. First of all, agonists

of dopamine receptors are used in Parkinson's disease (PD), which consists of denervation of striatal fibers projecting from the *substantia nigra*.¹ As an example, pramipexole is a non-ergot dopamine receptor agonist used in the therapy of PD and also of the restless legs syndrome.^{2–4}

The piperazinyl pyrimidine (**I**, Figure 1) is a privileged motif present in a number of therapeutic agents showing anti-neuroinflammatory activity, such as compound GIBH-130, able to suppress the IL-1 β in LPS-activated microglia, approved by China Food and Drug Administration for clinical

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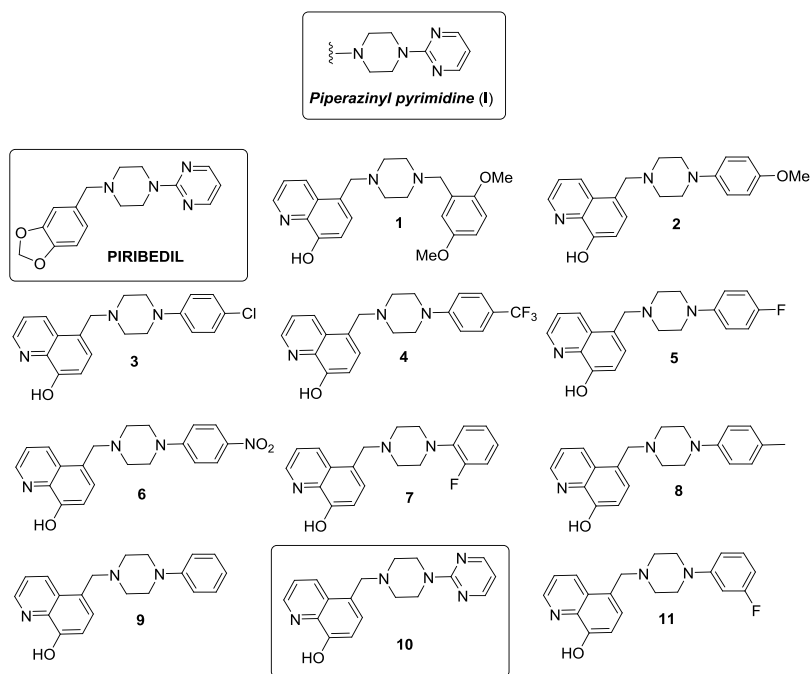


Figure 1. General structure of the piperaziny pyrimidine motif (I), pibredil, and the 5-((arylpiperazin-1-yl)methyl)quinolin-8-ols 1–11.

trials;⁵ MW01-070C, a novel anti-neuroinflammatory agent;⁶ Minozac, a drug under development as a water-soluble oral drug candidate for Alzheimer's disease;⁷ MW01-5-188WH, a new molecule that selectively inhibits glial activation in the CNS,⁸ or pibredil (Figure 1), an agent used for treating early and advanced PD that blocks α_2 -adrenoreceptors with minor effects on serotonergic, cholinergic, and histaminergic receptors.^{9–11} Furthermore, and of interest for the present work, pibredil is a dopamine receptor agonist that also has potential in the therapy of PD.^{12–14} However, pibredil taken by patients leads to side effects,¹⁵ thus limiting its therapeutic potential. Pibredil is a mixed D₂/D₃ compound. It should be noted that D₃ compounds have potential to combat some of the side effects of dopamine-based therapies in PD, namely dyskinesia, which correlates with increases in the expression levels of the D₃ receptor.^{16,17} Pibredil may therefore be a lead compound for development of D₃ receptor-selective compounds, as well as for gaining a better understanding of the function differences between D₂ and D₃ receptors.

With these ideas in mind, and continuing with our program on the synthesis and biological evaluation of *N*-arylpiperazine derivatives,^{18,19} we have designed *N*-aryl analogues of pibredil, in which we have also substituted the 5-methylbenzo[*d*][1,3]-dioxole motif by the 5-methylquinolin-8-ol group, resulting in 5-((arylpiperazin-1-yl)methyl)quinolin-8-ol type of compounds 1–11 (Figure 1), incorporating a diverse array of electron-donating or electron-withdrawing substituents at different positions in the aryl ring. These modifications should have positive consequences because of the biological and physicochemical properties that the 8-hydroxyquinoline group underscored by drugs bearing this structural motif.^{20,21} As a result from these studies, we have identified 5-((4-(pyrimidin-2-yl)piperazin-1-yl)methyl)quinolin-8-ol (10) (Figure 1) as a compound that exerts a profound bias in dopamine receptor signaling.

2. RESULTS AND DISCUSSION

2.1. Chemistry. The 5-((4-phenylpiperazin-1-yl)methyl)-8-hydroxyquinoline derivatives 1, 2, 3,²² 4–6, 7,²³ 8, 9,²³ 10, and 11 were synthesized in a “one-pot reaction”, by condensation of a solution of 5-(chloromethyl)-8-hydroxyquinoline hydrochloride²⁴ [or 2-(bromomethyl)-1,4-dimethoxybenzene¹⁸ in the case of compound 1] with the appropriate commercially available piperazine, in EtOH/tetrahydrofuran (THF) (1/1), during 15 min, at reflux. All compounds showed good analytical data in excellent agreement with their structure or the data described in the literature.

Inspection of the pharmacological data available for compounds 1–11 on public sources²⁵ revealed that, apart from pibredil, data are known only for 5-((4-phenylpiperazin-1-yl)methyl)quinolin-8-ol (9) (ChEMBL 1457644). Compound 9 was tested on a total of 52 assays for which IC₅₀ and EC₅₀ data were measured for 6 and 12 of them, respectively. Among them, of mention is the measured EC₅₀ value of 3763 nM (pEC₅₀ of 5.42) for streptokinase A. However, most surprisingly, even though molecule 9 shows close resemblance to pibredil and contains a phenyl piperaziny moiety, a known privileged motif among dopamine active ligands, it was not tested on any G protein-coupled receptor.

2.2. Biological Activity. The effect of the compounds synthesized on the D₂ and D₃ receptors was first tested in cAMP determination assays. As both D₂ and D₃ receptors are coupled to G_i, agonists lead to inhibition of the adenylyl cyclase. A total of 11 compounds at concentrations 200 nM and 1000 nM were tested in D₂- or D₃-receptor-expressing cells treated with forskolin, which leads to increases in cAMP. Any agonistic receptor-mediated effect will lead to a significant reduction in forskolin-induced cAMP levels. On the basis of this first (see Figure 2) screening, products 5, 6, 9, and 10 were selected for further studies. Dose-responses curves for cAMP and pERK1/2 assays are shown in Figure 3 in which it is shown that pibredil was the most potent compound. Raclopride, a nonselective D₂/D₃ receptor antagonist, was

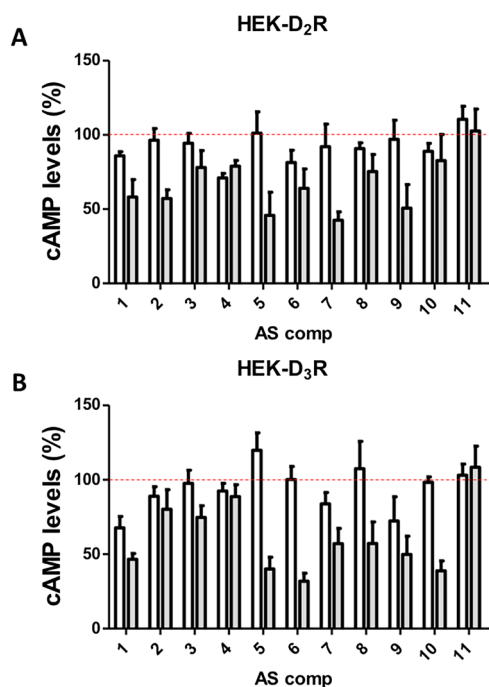


Figure 2. Signaling in HEK-293T cells expressing D₂R or D₃R. HEK-293T cells transfected with cDNA encoding for D₂R (A) or D₃R (B) were treated with compounds 1–11 (200 nM in white bars and 1000 nM in gray bars). cAMP accumulation was detected by time-resolved fluorescence energy transfer (TR-FRET) in the presence of 0.5 μ M forskolin. cAMP production is expressed as % of levels obtained by using 0.5 μ M forskolin. Data are mean \pm standard error of the mean (SEM) of three different experiments in triplicates.

able to revert the effect of piribedil and piribedil-related compounds (Figure 4). Furthermore, non-transfected cells did not respond to the dopamine receptor agonists (Figure S1).

For those selected compounds and for piribedil, dose–response effects were measured (Figure 3). This approach was performed in cAMP determination and ERK1/2 phosphorylation assays in both D₂- and D₃-receptor-expressing cells. IC₅₀ or EC₅₀ values and the percentage of response were calculated using sumanirole (100 nM) as selective D₂-receptor or 7-OH-PIPAT (100 nM) as selective D₃-receptor agonists. The results are summarized in Table 1 which presents IC₅₀ or EC₅₀ values plus the percentage of effect respect to a reference compound. As shown in Figure 4, the effect of compounds on either D₂- or D₃-receptor-expressing cells was blocked by 1 μ M raclopride, a potent nonselective D₂/D₃ receptor antagonist.

The results confirm that, in cAMP determination assays, piribedil is a preferential D₃ compound by 2 orders of magnitude.¹¹ However, a recent analysis of pharmacology data available for various compounds deposited in the ChEMBL database²⁵ revealed a fairly large data heterogeneity depending on the assay type and bioactivity endpoints.²⁶ Accordingly, we observed that piribedil selectivity for D₃ is lost when data on MAPK assays are considered. Interestingly, this loss of selectivity is very much in agreement with the known binding data available in public sources,²⁷ where the pK_i of piribedil for the D₂ and D₃ receptors is 6.85 and 6.60, respectively, albeit there is a difference of 3 orders of magnitude between binding pK_i data and our MAPK pEC₅₀ data. In contrast, the results for the compounds synthesized in this work prove that none displayed a clearly marked selectivity for any of the two receptors. Remarkably, the potency of the majority of the compounds was fairly high. However, the most salient finding was the differential effect when, compound by compound, cAMP and MAPK results were compared. Apart from the disappearance of the selectivity mentioned above of piribedil when comparing the EC₅₀ values in ERK1/2 phosphorylation assays, compound 10 showed a significant effect in cAMP assays but negligible capacity of activating the MAPK pathway.

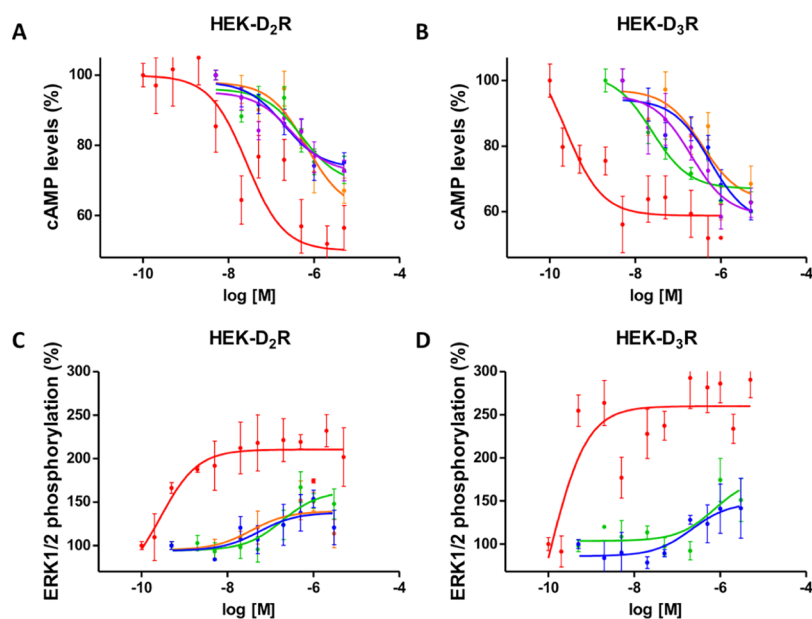


Figure 3. Dose response curves in HEK-293T cells expressing D₂R or D₃R. HEK-293T cells transfected with cDNA encoding for D₂R (A) or D₃R (B) were treated with piribedil (red) or compounds 5 (orange), 6 (green), 9 (blue), or 10 (purple). cAMP accumulation was detected by TR-FRET in the presence of 0.5 μ M forskolin. cAMP production is expressed as percentage of levels obtained by using 0.5 μ M forskolin. pERK1/2 phosphorylation is expressed as percentage over basal levels (100%). See General Methods for experimental details. Data are the mean \pm SEM of five different experiments in triplicates. The concentration range was 0.2–5000 nM for piribedil and 2–3000 nM for the rest of compounds.

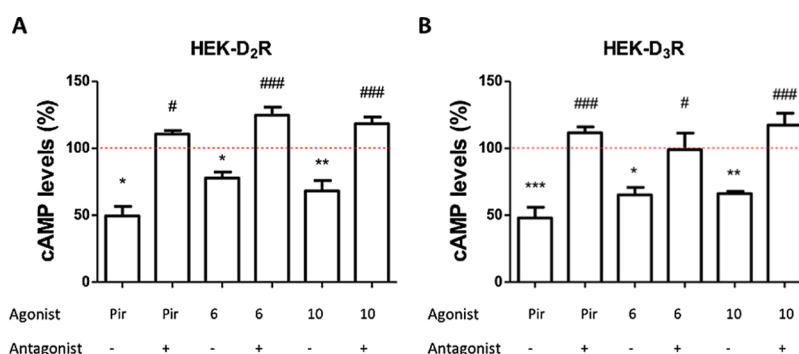
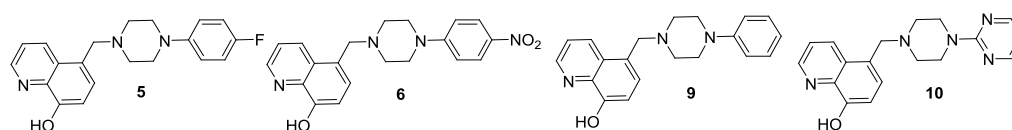


Figure 4. Reversion of agonist effect by raclopride in HEK-293T cells expressing D₂R or D₃R. HEK-293T cells transfected with cDNA encoding for D₂R (A, C) or D₃R (B, D) were pretreated with a dopamine receptor antagonist (1 μ M raclopride) and subsequently treated with piribedil or compounds **6** and **10** (300 nM). cAMP accumulation was detected by TR-FRET in the presence of 0.5 μ M forskolin. cAMP production is expressed as % of levels obtained by using 0.5 μ M forskolin. Data are the mean \pm SEM of three different experiments in triplicates. One-way ANOVA followed by Bonferroni's multiple comparison post-hoc test were used for statistical analysis. (* p < 0.05, ** p < 0.01, *** p < 0.001; vs treatment with forskolin); (# p < 0.05, ### p < 0.001; vs treatment with agonist).

Table 1. IC₅₀ (cAMP Assays), EC₅₀ (pERK1/2 MAPK Assays), and Percentage Values Versus Effect of 100 nM Sumanriole (D₂R) or 100 nM 7-OH-PIPAT (D₃R) of Piribedil and Compounds **5**, **6**, **9**, and **10**^a



compound	HEK-D ₂ R reference: 100 nM sumanriole		HEK-D ₃ R reference: 100 nM 7-OH-PIPAT	
	cAMP pIC ₅₀ \pm SD	MAPK pEC ₅₀ \pm SD	cAMP pIC ₅₀ \pm SD	MAPK pEC ₅₀ \pm SD
piribedil	7.5 \pm 0.5	9.5 \pm 0.4	9.6 \pm 0.6	9.5 \pm 0.9
	167%	131%	118%	179%
5	6.1 \pm 0.4	7.4 \pm 0.9	6.4 \pm 0.7	no effect
	129%	47%	105%	
6	6.2 \pm 0.5	6.7 \pm 0.4	7.7 \pm 0.5	6.3 \pm 1.3
	103%	75%	94%	50%
9	6.7 \pm 0.4	7.3 \pm 0.7	6.2 \pm 0.4	6.6 \pm 0.4
	88%	45%	125%	54%
10	6.5 \pm 0.6	no significant effect	6.7 \pm 0.5	no significant effect
	93%		118%	

^aPercentages respect the effect of D₂ or D₃ receptor reference compounds were calculated using the maximal effect obtained after data fitting. Compounds **5** and **10** did not led to any significant ERK1/2 phosphorylation.

Therefore, molecule **10** disclosed a biased signaling in both D₂ and D₃ receptors.

This is important, as novel drug discovery approaches include allosteric modulators and biased agonists.

3. CONCLUSIONS

To sum up, in this report, we have described the design, synthesis, and biological evaluation on D₂ or D₃ receptors of new piribedil derivatives. Our results show that the synthesized compounds bind to the orthosteric center and not to any allosteric site. Remarkably, molecule **10** shows a marked biased agonism in both D₂ and D₃ receptors. Whereas variation in intracellular cAMP levels is surely due to engagement of G_i, ERK phosphorylation likely results from G-protein-independent signaling. Activation of G proteins involves large-scale movements of the α -helical domain of the α -subunit of the heterotrimeric G protein; the movement is necessary for GDP dissociation.^{28–31} This mechanism of G protein activation occurs for products **5**, **6**, **9**, and **10** when they bind to and activate D₂R or D₃R (Table 1). The marked bias of piperazine

10 with absolute lack of action on the MAPK pathway is a most interesting, but unexpected result. Therefore, agent **10** can be considered a new tool compound to study the mechanisms of functional selectivity between D₂ and D₃ dopamine receptors, which are still poorly understood.

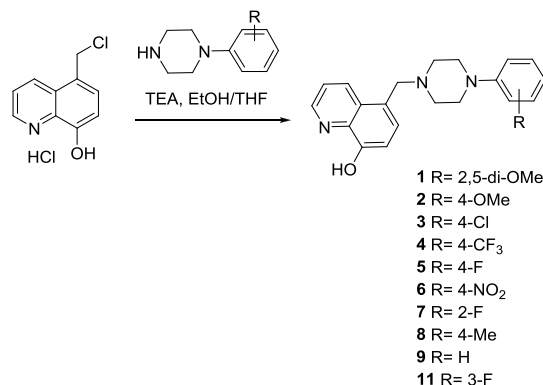
4. EXPERIMENTAL SECTION

4.1. Chemistry. Synthesis of Compounds 1–11.

4.1.1. General Methods. Melting points were determined in a Koffler apparatus and are uncorrected. ¹H NMR and ¹³C NMR spectra were recorded at room temperature in CDCl₃ or DMSO-*d*₆ at 400 MHz and at 100.6 MHz, respectively, using solvent peaks [CDCl₃: 7.26 (D), 77.2 (C) ppm and DMSO-*d*₆ 2.50 (D) and 39.7 (C) ppm] as internal references. The assignment of chemical shifts is based on standard NMR experiments (¹H, ¹³C, ¹H–¹H COSY, ¹H–¹³C HSQC, HMBC). Mass spectra were recorded on a gas chromatography/mass spectrometry (MS) spectrometer with an API-ES ionization source. Elemental analyses were performed at the IQOG (CSIC, Spain). Thin-layer chromatography analyses

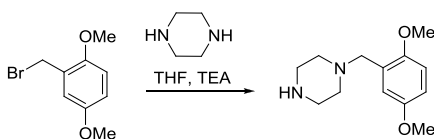
were performed on silica F254 and detection by UV light at 254 nm. Column chromatography was performed on silica gel 60 (230 mesh) (Scheme 1).

Scheme 1. Synthesis of 5-((Arylpiperazin-1-yl)methyl)quinolin-8-ol Derivatives 1–9 and 11



4.1.2. Synthesis of 5-((Arylpiperazin-1-yl)methyl)quinolin-8-ol Derivatives. To a solution of 1-arylpiperazine derivatives (0.1 mmol) in EtOH/THF (5/5 mL, v/v), TEA (three drops) and 5-(chloromethyl)quinolin-8-ol hydrochloride³⁰ (0.1 mmol) were added. The reaction was refluxed for the indicated time in each case. The solvent was evaporated, and the crude was diluted with CH₂Cl₂, washed with water, and extracted with CH₂Cl₂. The organic layer was dried with sodium sulfate and concentrated. Recrystallization in ethanol afforded the corresponding products as pure white solids (Scheme 2).

Scheme 2. Synthesis of 1-(2,5-Dimethoxybenzyl)piperazine



4.1.3. 1-(2,5-Dimethoxybenzyl)piperazine.³² To a solution of piperazine (0.516 g, 6 mmol) in dry THF (5 mL), a solution of 2-(bromomethyl)-1,4-dimethoxybenzene¹⁸ (0.231 g, 1 mmol) in THF (2 mL) was added dropwise. The reaction was refluxed for 30 min. Then, the reaction was diluted with EtOAc, washed with water, and extracted with EtOAc. The organic layer was dried with sodium sulfate, and the solvent was evaporated to afford a white solid 1-(2,5-dimethoxybenzyl)piperazine²² (0.242 g, 99%); R_f = 0.1 (hexane/EtOAc, 1/1, v/v); IR (KBr) ν : 3002, 2948, 2832, 1497 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 6.97 (dd, *J* = 3.0, 0.4 Hz, 1H), 6.77 (s, 1H), 6.74 (d, *J* = 3.0 Hz, 1H), 3.76 (s, 3H, OMe), 3.76 (s, 3H, OMe), 3.51 (s, 2H, CH₂), 2.89 (t, *J* = 4.9 Hz, 4H, 2× CH₂), 2.47 (s, 4H, 2CH₂), 2.24 (s, 1H, NH); ¹³C NMR (101 MHz, CDCl₃): δ 153.4, 152.0, 127.3, 116.3, 112.0, 111.5, 56.5, 56.1, 55.6, 54.2, 46.0; MS (EI⁺, *m/z*): 236.3 [M]⁺, 194.02, 151.1.

4.1.4. 5-((4-(2,5-Dimethoxybenzyl)piperazin-1-yl)methyl)quinolin-8-ol (1). (0.217 g, 67%); R_f = 0.3 (dichloromethane/methanol, 10/1); mp 145–6 °C; IR (KBr) ν : 3331, 3021, 2798, 2657, 1580, 1511 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.75 (dd, *J* = 4.2, 1.6 Hz, 1H), 8.65 (dd, *J* = 8.5, 1.6 Hz, 1H), 7.43 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.31 (d, *J* = 7.7 Hz, 1H), 7.05

(d, *J* = 7.7 Hz, 1H), 6.95 (d, *J* = 2.9 Hz, 1H), 6.73 (dt, *J* = 8.8, 5.9 Hz, 2H), 3.77 (s, 2H), 3.75 (s, 3H), 3.73 (s, 3H), 3.52 (s, 2H), 2.48 (s, 7H); ¹³C NMR (101 MHz, CDCl₃): δ 153.3, 151.9, 151.6, 147.4, 138.5, 134.1, 128.7, 127.8, 127.4, 124.7, 121.3, 116.3, 112.0, 111.4, 108.5, 60.5, 56.0, 55.8, 55.6, 53.1, 53.0; MS (ESI⁺, *m/z*): 394.2 [M + H]⁺. Anal. Calcd for C₂₃H₂₇N₃O₃: C, 70.21; H, 6.92; N, 10.68. Found: C, 70.44; H, 7.01; N, 10.77.

4.1.5. 5-((4-(4-Methoxyphenyl)piperazin-1-yl)methyl)quinolin-8-ol (2). (Reaction time: 15 min; 0.320 g, 92%); R_f = 0.5 (dichloromethane/methanol, 10/1); IR (KBr) ν : 3315, 2935, 2817, 2763, 1875, 1581, 1509, 1484 cm⁻¹; mp 173–4 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.77 (dd, *J* = 4.2, 1.5 Hz, 1H), 8.69 (dd, *J* = 8.5, 1.5 Hz, 1H), 7.45 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.35 (d, *J* = 7.7 Hz, 1H), 7.08 (d, *J* = 7.7 Hz, 1H), 6.90–6.77 (m, 4H), 3.84 (s, 2H, CH₂), 3.74 (s, 3H, OCH₃), 3.07–3.00 (m, 4H, 2× CH₂, piperazine), 2.67–2.57 (m, 4H, 2× CH₂, piperazine); ¹³C NMR (101 MHz, CDCl₃): δ 148.9, 147.0, 142.8, 140.9, 133.9, 129.4, 124.2, 123.1, 119.7, 116.6, 113.3, 109.5, 103.9, 55.9 (CH₂), 50.7 (OCH₃), 48.4 (2× CH₂, piperazine), 45.9 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 350.19 [M + H]⁺. Anal. Calcd for C₂₁H₂₃N₃O₂: C, 72.18; H, 6.63; N, 12.03. Found: C, 72.01; H, 6.53; N, 11.89.

4.1.6. 5-((4-(4-Chlorophenyl)piperazin-1-yl)methyl)quinolin-8-ol (3).²² (Reaction time: 15 min; 0.340 g, 96%); R_f = 0.7 (dichloromethane/methanol, 10/1); mp 191–2 °C; IR (KBr) ν : 3304, 2825, 2773, 1865, 1596, 1579, 1498, 1480 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.77 (dd, *J* = 4.2, 1.5 Hz, 1H), 8.68 (dd, *J* = 8.5, 1.6 Hz, 1H), 7.45 (dd, *J* = 8.6, 4.2 Hz, 1H), 7.35 (d, *J* = 7.7 Hz, 1H), 7.19–7.12 (m, 2H), 7.07 (d, *J* = 7.7 Hz, 1H), 6.88–6.75 (m, 2H), 3.84 (s, 2H, CH₂), 3.20–3.01 (m, 4H, 2× CH₂, piperazine), 2.70–2.50 (m, 4H, 2× CH₂, piperazine); ¹³C NMR (101 MHz, CDCl₃): δ 151.8, 149.9, 147.5, 138.6, 134.1, 129.0, 128.8, 127.8, 124.3, 124.2, 121.4, 117.1, 108.6, 60.6 (CH₂), 52.8 (2× CH₂, piperazine), 49.1 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 354.13 [M + H]⁺, 197.08, 158.07. Anal. Calcd for C₂₀H₂₀ClN₃O: C, 67.89; H, 5.70; N, 11.88. Found: C, 67.72; H, 5.88; N, 12.01.

4.1.7. 5-((4-(4-Trifluoromethyl)phenyl)piperazin-1-yl)methyl)quinolin-8-ol (2i = 4). (Reaction time: 15 min; 0.363 g, 94%); R_f = 0.6 (dichloromethane/methanol, 10/1); mp 163–4 °C; IR (KBr) ν : 3317, 2820, 2771, 1897, 1616, 1578, 1508 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.79 (dd, *J* = 4.2, 1.6 Hz, 1H), 8.70 (dd, *J* = 8.6, 1.6 Hz, 1H), 7.50–7.43 (m, 3H), 7.36 (d, *J* = 7.7 Hz, 1H), 7.10 (d, *J* = 7.7 Hz, 1H), 6.89 (d, *J* = 8.7 Hz, 2H), 3.86 (s, 2H), 3.30–3.14 (m, 4H), 2.65–2.56 (m, 4H); ¹³C NMR (101 MHz, CDCl₃): δ 153.2, 151.9, 147.6, 138.7, 134.1, 129.1, 127.8, 126.4, 126.3, 126.3, 126.3, 126.1, 124.1, 123.4, 121.5, 120.5, 120.1, 114.4, 108.6, 60.7 (CH₂), 52.7(2× CH₂, piperazine), 48.0 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 387.89 [M + H]⁺, 231.16, 157.82. Anal. Calcd for C₂₁H₂₀F₃N₃O: C, 65.11; H, 5.20; N, 10.85. Found: C, 65.32; H, 5.09; N, 10.77.

4.1.8. 5-((4-(4-Fluorophenyl)piperazin-1-yl)methyl)quinolin-8-ol (5). (Reaction time: 15 min; 0.302 g, 82%); R_f = 0.8 (dichloromethane/methanol, 10/1); mp 175–7 °C; IR (KBr) ν : 3297, 2938, 2821, 1869, 1580, 1505, 1477 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.79 (d, *J* = 3.5 Hz, 1H), 8.70 (d, *J* = 8.5 Hz, 1H), 7.47 (dd, *J* = 8.5, 4.1 Hz, 1H), 7.36 (d, *J* = 7.7 Hz, 1H), 7.09 (d, *J* = 7.7 Hz, 1H), 6.94 (dd, *J* = 11.9, 5.3 Hz, 2H), 6.89–6.79 (m, 2H), 3.86 (s, 2H), 3.13–3.02 (m, 4H), 2.68–2.55 (m, 4H); ¹³C NMR (101 MHz, CDCl₃): δ 158.2, 155.9, 151.9, 148.0, 148.0, 147.6, 138.7, 134.1, 129.0, 127.9,

124.3, 121.4, 117.7, 117.7, 115.5, 115.3, 108.6, 60.6 (CH₂), 53.0 (2× CH₂, piperazine), 50.18 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 338.21 [M + H]⁺, 180.95, 158.06. Anal. Calcd for C₂₀H₂₀FN₃O: C, 71.20; H, 5.98; N, 12.45. Found: C, 71.33; H, 5.77; N, 12.26.

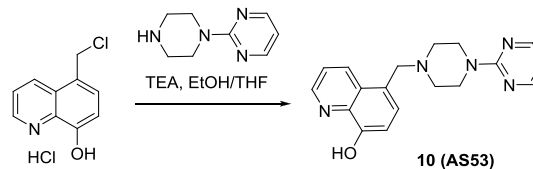
4.1.9. 5-((4-(4-Nitrophenyl)piperazin-1-yl)methyl)quinolin-8-ol (6). (Reaction time: 30 min; 0.320 g, 88%); R_f = 0.6 (dichloromethane/methanol, 10/1); mp 194–5 °C; IR (KBr) ν : 3392, 3006, 2934, 2833, 1837, 1587, 1503, 1459 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.78 (d, *J* = 4.1 Hz, 1H), 8.68–8.63 (m, 1H), 8.12–8.06 (m, 2H), 7.46 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.34 (d, *J* = 7.7 Hz, 1H), 7.08 (d, *J* = 7.7 Hz, 1H), 6.78 (t, *J* = 6.4 Hz, 2H), 3.85 (s, 2H, CH₂), 3.40–3.32 (m, 4H, 2× CH₂, piperazine), 2.63–2.55 (m, 4H, 2× CH₂, piperazine); ¹³C NMR (101 MHz, CDCl₃): δ 154.8, 152.0, 147.6, 138.6, 138.2, 134.0, 129.1, 127.8, 125.9, 123.8, 121.5, 112.5, 108.6, 60.5 (CH₂), 52.4 (2× CH₂, piperazine), 47.0 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 365.14 [M + H]⁺, 158.07. Anal. Calcd for C₂₀H₂₀N₄O₃: C, 65.92; H, 5.53; N, 15.38. Found: C, 65.71; H, 5.67; N, 15.44.

4.1.10. 5-((4-(2-Fluorophenyl)piperazin-1-yl)methyl)quinolin-8-ol (7).²³ (Reaction time: 15 min; 0.330 g, 98%); R_f = 0.6 (dichloromethane/methanol, 10/1); mp 186–8 °C; IR (KBr) ν : 3307, 2935, 2812, 1863, 1610, 1578, 1505, 1477 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.78 (dd, *J* = 4.2, 1.5 Hz, 1H), 8.75–8.60 (m, 1H), 7.46 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.36 (d, *J* = 7.7 Hz, 1H), 7.08 (d, *J* = 7.7 Hz, 1H), 7.05–6.99 (m, 2H), 6.98–6.86 (m, 2H), 3.86 (s, 2H), 3.04 (d, *J* = 4.2 Hz, 4H), 2.70–2.58 (m, 4H); ¹³C NMR (101 MHz, CDCl₃): δ 156.9, 154.4, 151.8, 147.5, 140.1, 140.1, 138.6, 134.1, 129.0, 127.9, 124.4, 124.4, 124.3, 122.3, 122.2, 121.34, 118.8, 118.8, 116.1, 115.9, 108.7, 60.6 (CH₂), 53.1 (2× CH₂, piperazine), 50.5 (CH₂, piperazine), 50.5 (CH₂, piperazine); MS (ESI⁺, *m/z*): 338.21 [M + H]⁺, 181.01, 158.13. Anal. Calcd for C₂₀H₂₀FN₃O: C, 71.20; H, 5.98; N, 12.45. Found: C, 71.09; H, 5.71; N, 12.63.

4.1.11. 5-((4-(*p*-Tolyl)piperazin-1-yl)methyl)quinolin-8-ol (8). (Reaction time: 15 min; 0.369 g, 98%); R_f = 0.5 (dichloromethane/methanol, 10/1); mp 167–8 °C; IR (KBr) ν : 3313, 2820, 2762, 1611, 1581, 1509, 1483 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.78 (dd, *J* = 4.2, 1.5 Hz, 1H, CH-2), 8.71 (dd, *J* = 8.6, 1.5 Hz, 1H, CH-4), 7.47 (dd, *J* = 8.6, 4.2 Hz, 1H, CH-3), 7.37 (d, *J* = 7.7 Hz, 1H), 7.07 (dd, *J* = 14.0, 8.1 Hz, 3H), 6.82 (d, *J* = 8.6 Hz, 2H), 3.86 (s, 2H, CH₂), 3.17–2.96 (m, 4H, 2× CH₂, piperazine), 2.70–2.56 (m, 4H, 2× CH₂, piperazine), 2.26 (s, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃): δ 151.8, 149.2, 147.6, 138.7, 134.1, 129.6, 129.1, 129.0, 127.9, 121.5, 116.4, 108.6, 60.7(CH₂), 53.1 (2× CH₂, piperazine), 49.7 (2× CH₂, piperazine), 20.4 (CH₃); MS (ESI⁺, *m/z*): 334.24 [M + H]⁺, 177.17, 158.17. Anal. Calcd for C₂₁H₂₃N₃O: C, 75.65; H, 6.95; N, 12.60. Found: C, 75.82; H, 6.75; N, 12.39.

4.1.12. 5-((4-(4-Phenylpiperazin-1-yl)methyl)quinolin-8-ol (9).²³ (Reaction time: 25 min; 0.430 g, 98%); R_f = 0.8 (dichloromethane/methanol, 10/1); mp 189–190 °C; IR (KBr) ν : 3312, 2816, 1599, 1578, 1505, 1493, 1478 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.80 (d, *J* = 4.2 Hz, 1H, CH-2), 8.72 (dd, *J* = 8.6, 1.3 Hz, 1H, CH-4), 7.48 (ddd, *J* = 8.5, 4.2, 0.6 Hz, 1H, CH-3), 7.38 (d, *J* = 7.7 Hz, 1H), 7.30–7.21 (m, 1H), 7.10 (d, *J* = 7.7 Hz, 1H), 6.92 (d, *J* = 8.5 Hz, 1H), 6.85 (td, *J* = 7.7, 0.6 Hz, 1H), 3.87 (s, 2H, CH₂), 3.20–3.13 (m, 4H, 2CH₂, piperazine), 2.69–2.59 (m, 4H, 2CH₂, piperazine); ¹³C NMR (101 MHz, CDCl₃): δ 147.0, 146.5, 142.8, 133.9,

129.4, 124.3, 124.2, 123.1, 119.6, 116.6, 114.8, 111.2, 103.8, 55.9 (CH₂), 48.2 (2× CH₂, piperazine), 44.4 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 320.19 [M + H]⁺, 161.08, 157.76. Anal. Calcd for C₂₀H₂₁N₃O: C, 75.21; H, 6.63; N, 13.16. Found: C, 75.42; H, 6.83; N, 13.09.



4.1.13. 5-((4-(Pyrimidin-2-yl)piperazin-1-yl)methyl)quinolin-8-ol (10). Prepared from 5-(chloromethyl)quinolin-8-ol hydrogen chloride,²⁴ and 2-(piperazin-1-yl)pyrimidine (reaction time: 30 min; 0.307 g, 96%); R_f = 0.7 (dichloromethane/methanol, 10/1); mp 143–5 °C; IR (KBr) ν : 3342, 2811, 1593, 1542, 1500, 1475 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.78 (dd, *J* = 4.2, 1.5 Hz, 1H), 8.70 (dd, *J* = 8.5, 1.5 Hz, 1H), 8.27 (dd, *J* = 4.7, 0.6 Hz, 2H), 7.46 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.33 (d, *J* = 7.7 Hz, 1H), 7.07 (d, *J* = 7.7 Hz, 1H), 6.45 (t, *J* = 4.7 Hz, 1H), 3.81 (s, 2H, CH₂), 3.78–3.73 (m, 4H, 2× CH₂, piperazine), 2.56–2.43 (m, 4H, 2× CH₂, piperazine); ¹³C NMR (101 MHz, DMSO-*d*₆): δ 161.5, 157.7, 151.8, 147.5, 138.7, 134.1, 129.0, 127.9, 124.3, 121.5, 109.7, 108.6, 60.8 (CH₂), 52.9 (2× CH₂, piperazine), 43.7 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 322.14 [M + H]⁺, 1665.19, 158.16. Anal. Calcd for C₁₈H₁₉N₅O: C, 67.27; H, 5.96; N, 21.79. Found: C, 67.41; H, 6.11; N, 21.64.

4.1.14. 5-((4-(3-Fluorophenyl)piperazin-1-yl)methyl)quinolin-8-ol (11). (Reaction time: 15 min; 0.279 g, 83%); R_f = 0.7 (hexane/EtOAc, 1/1); mp 162–3 °C; IR (KBr) ν : 3312, 2936, 2820, 1612, 1579, 1506 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.79 (dd, *J* = 4.2, 1.5 Hz, 1H), 8.70 (dd, *J* = 8.5, 1.5 Hz, 1H), 7.47 (dd, *J* = 8.5, 4.2 Hz, 1H), 7.36 (d, *J* = 7.7 Hz, 1H), 7.16 (dd, *J* = 15.3, 8.2 Hz, 1H), 7.09 (d, *J* = 7.7 Hz, 1H), 6.64 (dd, *J* = 8.4, 2.3 Hz, 1H), 6.53 (ddt, *J* = 15.3, 8.2, 2.4 Hz, 2H), 3.85 (s, 2H), 3.20–3.11 (m, 4H), 2.67–2.53 (m, 4H); ¹³C NMR (101 MHz, CDCl₃): δ 165.0, 162.6, 153.0, 152.9, 151.9, 147.6, 138.7, 134.1, 130.1, 130.0, 129.0, 127.9, 124.2, 121.5, 111.1, 111.0, 108.6, 105.8, 105.6, 102.7, 102.4, 60.6 (CH₂), 52.8 (2× CH₂, piperazine), 48.6 (2× CH₂, piperazine); MS (ESI⁺, *m/z*): 338.27 [M + H]⁺, 181.08, 158.06. Anal. Calcd for C₂₀H₂₀FN₃O: C, 71.20; H, 5.98; N, 12.45. Found: C, 71.36; H, 6.02; N, 12.02.

4.2. Biological Evaluation. **4.2.1. Cell Culture and Transfection.** Human embryonic kidney 293T (HEK-293T) cells were grown in Dulbecco's modified Eagle's medium supplemented with 2 mM L-glutamine, 1 mM sodium pyruvate, 100 units/mL penicillin/streptomycin, and 5% (v/v) fetal bovine serum [all supplements were from Invitrogen, (Paisley, Scotland, UK)]. Cells were maintained at 37 °C in a humidified atmosphere of 5% CO₂ and were passaged, with trypsin (Thermo Fisher Scientific, Waltham, MA, USA), when they were 80–90% confluent, that is, approximately twice a week. Cells were transiently transfected with the polyethyleneimine (Sigma, St. Louis, MO, USA) method as previously described.^{31,32} Experiments were carried out in cells expressing dopamine D₂ or D₃ receptors.

4.2.2. cAMP Determination and ERK Phosphorylation Assays. Details of the protocols are provided elsewhere.^{33,34} For cAMP assays, cells were serum deprived and a phosphodiesterases inhibitor was used (50 μM zardaverine).

Agonists were added 15 min before addition of forskolin (0.5 μ M) forskolin, and reaction took place for 60 min (25 °C). The cAMP level was determined using the Lance Ultra cAMP kit (PerkinElmer, Waltham, MA, USA), and readings were performed in a PHERAstar (BMG Lab technologies, Offenburg, GE) equipped with an HTRF optical module. For pERK1/2 determination, the procedure was similar, but cells were washed prior use of lysing buffer (20 min 0°–4°). Supernatants (10 μ L) were analyzed using ad hoc AlphaScreen SureFire and the EnSpire Reader (both from PerkinElmer, Waltham, MA, USA). When indicated, the antagonist was added 15 min prior to the addition of the agonist. The lack of effect (cAMP or pERK1/2 assays) in non-transfected cells is shown in Supporting Information (Figure S1).

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.8b02509.

Lack of effect of piribedil and compounds **6** or **10** in cells that do not express dopamine receptors, that is, in untransfected HEK-293T cells (Figure S1) (PDF)

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Notes

The authors declare no competing financial interest.

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