

1 **ROLE OF CANNABINOIDS IN ALCOHOL-INDUCED**
2 **NEUROINFLAMMATION**

3
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27 Abbreviations

AEA	N-arachidonoylethanolamine or anandamide
AP-1	Activator protein 1
BBB	Blood-brain barrier
BDNF	Brain-derived neurotrophic factor
cAMP	Cyclic adenosine monophosphate
CB1	Cannabinoid receptor 1
CB2	Cannabinoid receptor 2
CBD	Cannabidiol
CBDA	Cannabidiolic acid
CBG	Cannabigerol
CBGV	Cannabigivarin
CNS	Central nervous system
COX-2	Cyclooxygenase-2
DAGL	Diacylglycerol lipase
DAMPs	Danger associated molecular patterns
eCB	Endocannabinoid
ECS	Endocannabinoid system
ERK	Extracellular signal-regulated kinase
FAAH	Fatty acid amide hydrolase
GFAP	Glial fibrillary acidic protein
GPCR	G protein-coupled receptor
HMGB1	High mobility group box 1
HPC	Hippocampus
Iba1	Ionized calcium binding adaptor molecule 1
IL	Interleukin
INF- γ	Interferon gamma
iNOS	Inducible nitric oxide synthase
I κ B α	Inhibitory kappa B α
LPS	Lipopolysaccharide
MAGL	Monoacylglycerol lipase
MCP-1	Monocyte chemoattractant protein 1
MCSF	Macrophage colony-stimulating factor
MD2	Myeloid differentiation protein-2
MHCII	Major histocompatibility complex II
MIP-1 α	Macrophage inflammatory protein 1 α
miRNA	MicroRNA
MRF-1	Microglia response factor 1
MyD88	Myeloid differentiation factor 88
NAPE-PDL	N-acylphosphatidylethanolamine-specific phospholipase d-like hydrolase
Nrf2	Nuclear factor erythroid 2-related factor 2
NF- κ B	Nuclear factor-kappa B
OEA	Oleylethanolamide

PFC	Prefrontal cortex
PKA	Protein kinase A
PPAR	Proliferator-activated receptor
ROS	Reactive oxygen species
RXR- γ	Retinoid X receptor- γ
SDF-1 α /CXCL1	Chemokine stromal cell-derived factor 1
SOC	Suppressors of cytokine signalling
TAK1	Transforming growth factor beta-activated kinase 1
TFG	Transforming growth factor
THC	Tetrahydrocannabinol
THCA	Tetrahydrocannabinolic acid
THCV	Tetrahydrocannabivarin
TLR4	Toll-like receptor 4
TNF- α	Tumour necrosis factor alpha
TrkB	Tropomyosin receptor kinase B
TRPV	Transient receptor potential vanilloid type

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29

30 **Highlights**

- 31 • Alcohol exposure leads to a chronic pro-inflammatory profile in the CNS.
- 32 • The expanded endocannabinoid system is a key regulator of the neuroimmune
33 response.
- 34 • CB1 activation is related to deterioration of alcohol-induced neuroinflammation.
- 35 • Cannabinoids modulate the pro-inflammatory and the anti-inflammatory
36 signalling pathways.
- 37 • Targeting the PPAR γ or the usage of OEA leads to promising outcomes.

38 **Abstract**

39 Alcohol is a psychoactive substance highly used worldwide, whose harmful use might
40 cause a broad range of mental and behavioural disorders. Underlying brain impact, the
41 neuroinflammatory response induced by alcohol is recognised as a key contributing factor
42 in the progression of other neuropathological processes, such as neurodegeneration.
43 These sequels are determined by multiple factors, including age of exposure. Strikingly,
44 it seems that the endocannabinoid system modulation could regulate the alcohol-induced
45 neuroinflammation. Although direct CB1 activation can worsen alcohol consequences,
46 targeting other components of the expanded endocannabinoid system may counterbalance
47 the pro-inflammatory response. Indeed, specific modulations of the expanded
48 endocannabinoid system have been proved to exert anti-inflammatory effects, primarily
49 through the CB2 and PPAR γ signalling. Among them, some endo- and exogenous
50 cannabinoids can block certain pro-inflammatory mediators, such as NF- κ B, thereby
51 neutralizing the neuroinflammatory intracellular cascades. Furthermore, a number of
52 cannabinoids are able to activate complementary anti-inflammatory pathways, which are
53 necessary for the transition from chronically overactivated microglia to a regenerative
54 microglial phenotype. Thus, cannabinoid modulation provides cooperative anti-
55 inflammatory mechanisms that may be advantageous to resolve a pathological
56 neuroinflammation in an alcohol-dependent context.

57 **1. Introduction**

58 Alcohol is one of the most consumed psychoactive drugs worldwide, with 2.3 billion
59 ocurrent drinkers. Indeed, alcohol use is among the leading causes of global disease
60 burden (Rehm and Shield, 2019). Around 3 million deaths per year occur due to the
61 harmful use of alcohol, which represents 5.3% of total deaths (World Health
62 Organization, 2018). Alcohol abuse affects most body organs, although severe alcohol-
63 induced diseases are most notable in the liver, pancreas and brain. This is due to alcohol's
64 direct neurotoxic effect on neurons through different mechanisms, including oxidative
65 stress (Qin and Crews, 2012) and neuroinflammation (Crews and Vetreno, 2014).

66 Even though alcohol exposure may have a different impact in the central nervous system
67 (CNS) determined by several factors, such as the developmental period of exposure, route
68 of administration or pattern of consumption, there might be a common
69 neuroinflammatory mechanism underlying such alterations. The endocannabinoid system
70 (ECS) has been observed to play a key role in the modulation of neuroinflammatory
71 responses (Chiurchiù et al., 2015; Cristino et al., 2020). In this context, we review and
72 discuss the role of cannabinoids in the regulation of alcohol-induced neuroinflammation.

73 In this review, we first outline the mechanisms by which alcohol exposure induces a
74 neuroinflammatory response. We also provide an overview on the ECS and its role in
75 regards with the regulation of neuroinflammation. Then, we discuss the molecular
76 mechanisms by which endocannabinoids, phytocannabinoids and synthetic cannabinoids
77 may regulate alcohol-induced neuroinflammatory responses. Additionally, we are
78 interested in alcohol impact on different neurobehavioral outcomes focused on preclinical
79 studies, using cannabinoid-based approaches to modulate the alcohol-induced brain
80 damage (i.e. neuroinflammation, oxidative damage or neurodegeneration) in two different
81 developmental stages: adolescence and adulthood. Finally, we propose a mechanism by
82 which cannabinoid-based therapies could modulate alcohol-induced neuroinflammatory
83 signalling.

84 **2. Alcohol-induced neuroinflammatory response.**

85 The immune response within the CNS involves peripheral and local elements, primarily
86 microglia and astrocytes, together forming the so-called glia (Gilhus and Deuschl, 2019).

87 Both cell types are highly reactive to changes in the brain, actively producing signalling
88 molecules that can either participate in homeostasis or contribute to disease if an insult is
89 present (Greenhalgh et al., 2020). Evidence suggests the existence of crosstalk between
90 activated microglia and astrocytes that amplifies the inflammatory response, contributing
91 to the production of neurotoxic factors (Saijo and Glass, 2011).

92 *2.1. Evidence from clinical studies*

93 Clinical studies have shown an upregulation of immune-related genes in post-mortem
94 brains of alcoholic patients (Lewohl et al., 2000). Increased protein levels of Monocyte
95 chemoattractant protein 1 (MCP-1) in the ventral tegmental area, substantia nigra,
96 hippocampus (HPC) and amygdala have been found in alcoholic post-mortem brains as
97 compared with controls (He and Crews, 2008). In addition, enriched expression of genes
98 associated with interferon signalling pathway in prefrontal cortex (PFC) has been shown
99 in alcohol use disorder (AUD) post-mortem brains (Kapoor et al., 2019).

100 Other clinical studies have shown changes in pro-inflammatory cytokines levels in
101 plasma of current alcohol drinkers. Circulating levels of interleukin (IL)-6 and tumor
102 necrosis factor alpha (TNF- α) were increased on the first day of withdrawal in alcohol-
103 dependent patients without cirrhotic liver disease (Heberlein et al., 2014), suggesting a
104 direct induction of pro-inflammatory cytokines in blood by alcohol. By contrast, other
105 authors highlighted the importance of alcohol-induced secondary disorders to promote
106 the enhancement of circulating inflammatory molecules in alcoholic patients (Achur et
107 al., 2010; González-Reimers et al., 2011). Additionally, at least one clinical study has
108 pointed out that intestinal permeability and lipopolysaccharide (LPS) circulating levels
109 were largely increased in alcohol-dependent patients at the onset of withdrawal, which
110 positively correlated with systemic pro-inflammatory cytokines. Interestingly, pro-
111 inflammatory cytokines (TNF- α , IL-6, and C-reactive protein) remained increased after
112 3 weeks of withdrawal (Leclercq et al., 2012).

113 *2.2. Evidence from preclinical studies*

114 In accordance with clinical data, some preclinical studies have displayed an increase of
115 pro-inflammatory markers after alcohol exposure even though several factors can
116 influence the alcohol's effect on neuroimmune activation. Vallés and colleagues

117 demonstrated an enhanced expression of inducible nitric oxide synthase (iNOS),
118 cyclooxygenase-2 (COX-2) and IL-1 β , within the cerebral cortex of female Wistar rats,
119 following a 5-months chronic alcohol treatment (Vallés et al., 2004). 10-days of alcohol
120 treatment (5 g/kg; i.g.) increased brain TNF- α and MCP-1 levels, whereas it reduced IL-
121 10, which is an anti-inflammatory cytokine, in adult male C57BL/6 mice (Qin et al.,
122 2008).

123 Other authors have suggested an age- and region-specific susceptibility to alcohol
124 regulation of neuroinflammatory response (Kane et al., 2014; Pascual et al., 2007; Perkins
125 et al., 2019). Although divergences could also be due to the usage of different models of
126 alcohol exposure, highlighting the importance of dose, route of administration and timing
127 of exposure in the alcohol-induced neuroinflammatory response. Nevertheless, aging-
128 associated changes in neuroimmune response are known to play a key role in adolescent
129 *versus* adult alcohol-induced neuroinflammation, which are to take into account (Perkins
130 et al., 2019). Hence, the period of lifetime in which alcohol exposure occurs is important
131 in terms of the severity of neuroinflammatory response and the following negative
132 consequences.

133 Sex factor seems to be relevant as well. When male and female mice were compared,
134 iNOS, COX-2, IL-1 β and TNF- α were increased in cerebral cortex of both sexes.
135 However, alcohol induced higher glial fibrillary acidic protein (GFAP) levels in females
136 *versus* males (Alfonso-Loeches et al., 2013). On the other hand, concerning the
137 methodology, the time point examining post-exposure, the techniques to analyse the
138 protein and mRNA content, and the species used can also influence the changes on
139 immune signature (see (Melbourne et al., 2019) for more information).

140 Thus, the interaction between alcohol and neuroimmune system is complex and still
141 remains unclear. Although the number of studies focused on this field has been increasing
142 in the last decades, reports are not particularly consistent because of the influence of many
143 external factors.

144 *2.3. Mechanisms of alcohol-induced neuroinflammation*

145 *2.3.1. Central mechanisms*

146 Taken together, it seems that alcohol exposure can be defined as an insult that leads to
147 neuroimmune dysregulation in some situations. These neuroimmune alterations are
148 mostly mediated by central mechanisms that lead to the activation of microglia, which
149 can persist for long periods once activated by excessive alcohol consumption (Liu et al.,
150 2008; Vetreno et al., 2013). Alcohol molecule can cross the blood-brain barrier (BBB) to
151 induce a local pro-inflammatory response due to the activation of Toll-like receptors
152 (TLRs) in glial cells (Coleman and Crews, 2018; Crews and Vetreno, 2016). In addition,
153 alcohol exposure may induce the release of endogenous danger associated molecular
154 patterns (DAMPs), such as the high mobility group box 1 (HMGB1), by neurons and glia
155 during glutamate hyperexcitation (Maroso et al., 2011; Zou and Crews, 2005) or by
156 necrotic dead cells (Crews and Vetreno, 2016). DAMPs in extracellular space activate
157 TLRs and other pro-inflammatory-associated receptors, such as the receptor for advanced
158 glycation end products (Park et al., 2004). The activation of these receptors induces the
159 stimulation of pro-inflammatory signalling pathways and activation of transcriptional
160 factors, such as nuclear factor kappa B (NF- κ B) and activator protein 1 (AP-1), through
161 the myeloid differentiation factor 88 (Myd88)-dependent pathway. Furthermore, a
162 hyperactivation of pro-inflammatory oxidases occurs when alcohol is metabolized,
163 resulting in the formation of reactive oxygen species (ROS). Altogether cause positive
164 loops of pro-inflammatory responses in the brain that converge upon NF- κ B activation
165 and its subsequent translocation into the nucleus, which will keep on amplifying the
166 neuroimmune gene induction through autocrine and paracrine positive feedbacks (Crews
167 and Vetreno, 2016).

168 During such periods of activation, microglia suffers morphological and functional
169 changes. Thus, authors suggested that a single period of binge alcohol drinking does not
170 induce a full microglial-driven neuroinflammatory response, but a partial microglial
171 activation that persists until adulthood (McClain et al., 2011). In a chronic but moderate
172 alcohol exposure model using a vapor chamber for 5 weeks, increased number of Iba1-
173 positive cells were observed at the first day of withdrawal in many brain regions,
174 including frontal cortex, HPC, amygdala, substantia nigra and cerebellum. Furthermore,
175 Iba1 staining remained elevated after 28 days of withdrawal in amygdala, frontal cortex
176 and substantia nigra (Sanchez-Alavez et al., 2019). Accordingly, a chronic alcohol
177 drinking model over 6 months induced long-lasting partial microglial activation in rat
178 HPC (Cruz et al., 2017). On the other hand, other authors have shown an increased

179 number of MHCII-, CD45- and CD68-positive cells in a chronic alcohol-drinking model
180 over 5 months in mice, suggesting an activated ameboid phenotype in this case. Elevated
181 levels of pro-inflammatory cytokines and chemokines, including chemokine ligand 2 and
182 fractalkine, were reported in this study following a chronic alcohol exposure (Alfonso-
183 Loeches et al., 2016).

184 Therefore, one may confirm that there is no clear consensus about the activated microglial
185 phenotype induced by alcohol and its functional implications. Again, there are several
186 factors that might lead to a different neuroinflammatory response. Interestingly, evidence
187 suggests a possible association between chronic or excessive alcohol consumption with
188 pro-inflammatory microglial phenotypes, whereas single or moderate drinking episodes
189 could be accompanied by homeostatic responses. However, further studies should be
190 addressed to clarify the influence of the factors mentioned upon the microglial phenotype,
191 alongside the role of this microglial activation following alcohol exposure. Thus, whether
192 this neuroimmune activation is detrimental or neuroprotective remains a subject of
193 debate.

194 However, some studies in attempt to assess causality have exhibited that the deletion of
195 TLR4 prevents the neurotoxicity through the blockade of alcohol-induced activation of
196 microglia (Erickson et al., 2019). TLR4 deletion prevented alcohol-induced upregulation
197 of CD11b (microglial marker) and GFAP immunoreactivity. Furthermore, in this
198 knockout mouse model the increase of caspase-3 and iNOS activity, as well as the
199 increase of COX-2, IL-1 β , TNF- α and IL-6 in the cerebral cortex of female mice, were
200 also prevented following a 5-month chronic alcohol treatment (Alfonso-Loeches et al.,
201 2010). Another study has shown that the inflammatory response induced by alcohol
202 treatment was completely abolished in microglia of TLR4-deficient mice (Fernandez-
203 Lizarbe et al., 2009). Altogether, it is relatively clear that the activation of microglia under
204 certain alcohol exposure conditions could be somehow detrimental for the CNS,
205 supporting the hypothesis of neuroimmune activation in the pathophysiology of AUDs.
206 Noteworthy, microglia may as well promote repair under many homeostatic and
207 pathological conditions, including other certain types of alcohol exposure mentioned
208 above.

209 *2.3.2. Peripheral mechanisms*

210 The presence of alcohol increases blood's innate immune signals released from peripheral
211 organs. The gut and liver are especially affected by alcohol intake as it has been reported
212 in different studies; therefore, pro-inflammatory markers can be released when they are
213 dramatically affected by alcohol exposure (de Timary et al., 2017; Ferrier et al., 2006;
214 Gao et al., 2011). Alcohol increases the intestinal permeability (Antón et al., 2018),
215 allowing the release of pathogen-associated molecular patterns into the bloodstream, such
216 as LPS endotoxin, stimulating pro-inflammatory cytokine production. These circulating
217 immune signals, primarily cytokines and chemokines (i.e.:TNF- α , IL-6, IL-1 β and
218 chemokine stromal cell-derived factor 1 (SDF-1 α /CXCL1)), are proposed to contribute
219 to neuroimmune activation in AUD as various immune-to-brain communication
220 pathways have been described (Banks and Erickson, 2010; Crews et al., 2006; de Timary
221 et al., 2017). First, circulating pro-inflammatory cytokines may activate the
222 hypothalamus-pituitary-adrenal axis through the vagus nerve stimulation (so-called
223 'neural pathway'). Moreover, peripheral signals can reach the brain via circumventricular
224 organs, meninges and the choroid plexus, structures with a 'leaky' BBB, and they can
225 also act on brain endothelium to induce the release of secondary mediators eliciting a
226 response into the brain (named 'humoral pathway'). In addition, endothelial cells may
227 become activated by circulating cytokines and secrete inflammatory mediators that alter
228 the permeability of the BBB, allowing the recruitment of peripheral immune cells (known
229 as 'cellular pathway') (Capuron and Miller, 2011; D'Mello and Swain, 2017; Dantzer et
230 al., 2008).

231 **3. Cannabinoids and neuroinflammation**

232 *3.1. The endocannabinoid system*

233 The search for specific binding sites for tetrahydrocannabinol (THC) lead to the first
234 identification of cannabinoid receptor 1 (CB1) (Devane et al., 1988) and cannabinoid
235 receptor 2 (CB2) (Matsuda et al., 1990; Munro et al., 1993). CB1 is considered to be the
236 most abundant G-protein coupled receptor (GPCR) within the CNS (Irving et al., 2008)
237 being primarily expressed in axon terminals (Freund et al., 2003). When activated, CB1
238 inhibits neurotransmitter release from both presynaptic glutamatergic (Katona et al.,
239 2006) and GABAergic neurons (Katona et al., 1999). CB2 is found in lower levels in the
240 brain, being mostly located in microglia (Atwood and Mackie, 2010; Walter et al., 2003).

241 The spread localization among immune cells confers CB2 a key role in the modulation of
242 inflammatory processes.

243 The most studied endogenous ligands to CB1 and CB2 are N-arachidonylethanolamine
244 or anandamide (AEA) and 2-arachidonoylglycerol (2-AG) (Lu and MacKie, 2016).
245 Endocannabinoids (eCB) are synthesized on demand and, unlike classic
246 neurotransmitters, are not stored into vesicles, but immediately released from
247 postsynaptic cells (Di Marzo et al., 1998). The main enzymes involved in the synthesis
248 of eCB are N-acylphosphatidylethanolamine-specific phospholipase d-like hydrolase
249 (NAPE-PLD) (Okamoto et al., 2004) and diacylglycerol lipase (DAGL) α and β (Bisogno
250 et al., 2003), for AEA and 2-AG respectively. The canonical metabolic pathway for eCBs
251 degradation is the hydrolysis of AEA by fatty acid amide hydrolase (FAAH) (Cravatt et
252 al., 1996) and 2-AG by monoacylglycerol lipase (MAGL) (Dinh et al., 2002). However,
253 eCBs can also be degraded by oxygenation via cyclooxygenase-2 (COX-2), among others
254 (Di Marzo et al., 2000).

255 These receptors, ligands and enzymes responsible for their synthesis and degradation
256 constitute the ECS, which is a neuromodulatory system engaged in a wide range of
257 physiological roles, such as brain development, homeostasis, neurotransmitter release,
258 synaptic plasticity and immune response (Lu and MacKie, 2016). In order to develop its
259 function, the ECS directly or indirectly interplays with other components of the CNS
260 leading to what some authors have called the expanded endocannabinoid system (Cristino
261 et al., 2020). Therefore, other N-acylethanolamines, such as oleoylethanolamide (OEA),
262 or 2-acylglycerols molecules, as well as long-chain N-acyl-amides can also interact with
263 cannabinoid and cannabinoid-like receptors, such as peroxisome proliferator-activated
264 receptor (PPAR), orphan receptor GPR or transient receptor potential (TRP) channel
265 (Cristino et al., 2020).

266 *3.2. Cannabinoid signalling in neuroinflammation*

267 The wide distribution of ECS throughout the CNS and the immune system confers
268 cannabinoids a privileged position for the regulation of neuroinflammatory responses.
269 Although CB1 is most expressed in neurons, it can also be found in glial cells in the brain
270 (Stella, 2010). The stimulation of CB1 has been shown to diminish the release of pro-
271 inflammatory cytokines, iNOS and ROS via NF- κ B pathway inhibition (Lou et al., 2016;

272 Ribeiro et al., 2013) and protect from excitotoxicity (Marsicano, 2003). Besides, some
273 studies have also proved that CB1 antagonists can indirectly induce an anti-inflammatory
274 response (Kaplan, 2013).

275 Due to its microglial localization, CB2 is highly involved in the modulation of
276 neuroimmune responses. CB2 levels in the CNS increase drastically in neurodegenerative
277 disorders (Aymerich et al., 2018; Cassano et al., 2017) or after brain insults (Cabral and
278 Griffin-Thomas, 2009). Nevertheless, the range of the alcohol-induced
279 neuroinflammatory response is probably less profound and so, its effects on central CB2
280 levels might be fewer. Still, CNR2 gene expression is increased in alcohol users and in
281 human monocyte-derived dendritic cells treated with alcohol (Agudelo et al., 2013),
282 although more studies focusing on the alcohol effects on central CB2 levels would be
283 clarifying. In fact, CB2 levels in microglia are phenotype-dependent, being principally
284 expressed in activated and primed microglia (Stella, 2010). The activation of CB2 is
285 related to decreases in pro-inflammatory cytokines (TNF- α , interferon gamma (IFN- γ),
286 IL-1, IL-2, IL-6 or IL-12) (Croxford and Yamamura, 2005; Mecha et al., 2016; Yuan et
287 al., 2002), chemokines (Bátkai et al., 2012; Sheng et al., 2005) and iNOS (Wen et al.,
288 2015; Zarruk et al., 2012) via inhibition of the NF- κ B pathway (Fakhfoury et al., 2012;
289 Jeon et al., 1996). Hence, CB2 would act as a homeostatic regulator, bringing the system
290 back to physiological states.

291 Cannabinoids also bind to other non-cannabinoid receptors in the so-called expanded
292 cannabinoid system in order to exert its anti-inflammatory and homeostatic functions.
293 eCBs (AEA and other endocannabinoid-like mediators) (Cristino et al., 2020),
294 phytocannabinoids (cannabidiol (CBD), cannabidiolic acid (CBDA), cannabigerol
295 (CBG), tetrahydrocannabinolic acid (THCA)) (Di Marzo, 2018) and synthetic
296 cannabinoids (WIN55,212-2) (Fakhfoury et al., 2012) are agonist of PPARs, which are
297 nuclear receptors that inhibit NF- κ B and AP1-mediated inflammation (Varga et al.,
298 2011a). In the CNS, PPARs are expressed in neurons and glial cells (Moreno et al., 2004).
299 Out of its three isoforms, PPAR α and specially PPAR γ are involved in the regulation of
300 neuroinflammation and lipid metabolism, whereas PPAR β/δ remains the least studied one
301 (Varga et al., 2011a). Interestingly, their activation has been shown to exert protective
302 effects in different alcohol intake models (Alen et al., 2018; Blednov et al., 2015;
303 Cippitelli et al., 2017).

304 TRP channels are highly related to the eCB signalling. Indeed, TRP vanilloid 1 (TRPV1),
305 TRPV2, TRPV3, TRPV4, TRP ankyrin 1 and TRP melastatin 8 have all been reported to
306 mediate cannabinoid activity (Muller et al., 2018). TRPV1 has recently been shown to
307 control microglial activation and glutamate release from microglial microvesicles
308 (Marrone et al., 2017). Another recent study also found that TRPV1 regulate cytokine
309 release from activated microglia (Bassi et al., 2019). The eCBs AEA and 2-AG (Lowin
310 and Straub, 2015; Petrosino et al., 2016), some phytocannabinoids, such as
311 tetrahydrocannabivarin (THCV), CBD, CBG or cannabigivarin (CBGV) (De Petrocellis
312 et al., 2011), and several synthetic cannabinoids, such as WIN55,212-2 (Soethoudt et al.,
313 2017), bind to TRPV1.

314 The orphan receptor GPR55 has also been proposed as an eCB receptor. Some studies
315 have revealed that AEA and 2-AG bind and activate GPR55 (Lauckner et al., 2008;
316 Ryberg et al., 2007), whereas CBD acts as an antagonist (Kaplan et al., 2017) and THC
317 as an agonist (Lauckner et al., 2008). Furthermore, atypical synthetic cannabinoids, other
318 than WIN55,212-2, can also activate this receptor (Johns et al., 2007). GPR55 levels are
319 microglia-activation dependent, mimicking the regulatory pattern of CB2 and conferring
320 a potential role in the regulation of inflammatory responses (Pietr et al., 2009). Other
321 orphan receptors could also play an important role in neuroinflammation: GPR18 due to
322 its microglial localization (McHugh, 2012), or GPR119 due to the binding of OEA,
323 among other cannabinoid-like mediators (Hansen et al., 2012). Nevertheless, little is
324 known about the physiological roles of these receptors as wells as their interactions with
325 the endocannabinoid system and the immune response which are yet to be elucidated.

326 Besides, due to their lipidic nature, eCBs are highly related to the eicosanoid system
327 (Grabner et al., 2017). Prostaglandins, a classical eicosanoid, are arachidonic acid-derived
328 bioactive lipid mediators synthetized by COX-1/2 that have a prominent role in the
329 regulation of inflammatory processes (Aoki and Narumiya, 2012; Dennis and Norris,
330 2015). Indeed, prostaglandins are thought to mediate the transition to chronic
331 inflammation via different mechanisms including the amplification of pro-inflammatory
332 cytokines (Chiurchiù et al., 2018). Although phospholipase A2 is considered to be the
333 main source of the prostaglandin precursor arachidonic acid, the hydrolysis of 2-AG by
334 MAGL has also been proved to generate it (Grabner et al., 2017; Long et al., 2009;
335 Schlosburg et al., 2010). In this line, MAGL inhibition might protect from the

336 prostaglandin-induced neuroinflammation, as evidenced in a mouse model of Parkinson
337 disease (Nomura et al., 2011).

338 The neuroimmune modulatory profile of cannabinoids has led to the emergence of a wide
339 range of studies exploiting these molecules for the treatment of neurological and
340 neurodegenerative diseases. Endogenous, phytogenic and synthetic cannabinoids have
341 been proved to ameliorate neuroinflammation induced by different pathologies, such as
342 Alzheimer's disease (Maroof et al., 2013; Vallée et al., 2017), Parkinson's disease (Kelly
343 et al., 2020; Little et al., 2011), multiple sclerosis (Al-Ghezi et al., 2019; Correa et al.,
344 2007), neuropathic pain (Donvito et al., 2018), autism spectrum disorder (Araujo et al.,
345 2019) or addiction (Rodrigues et al., 2014).

346 **4. Cannabinoids in alcohol-induced neuroinflammation**

347 Alcohol exposure may have a different impact on neurobehavioral outcomes depending
348 on the developmental timing of exposure. Subsequently, we divided our discussion about
349 the role of cannabinoids in alcohol-induced neuroinflammation by the developmental
350 period at which alcohol exposure took place.

351 *4.1. Adolescent alcohol exposure*

352 Adolescence represents a period in which the brain is undergoing extensive maturational
353 processes. Therefore, alcohol consumption during this time can cause structural and
354 functional changes in immature brain areas resulting in cognitive and behavioural deficits
355 (Kyzar et al., 2016). Compelling evidence has shown that the activation of the immune
356 system plays a crucial role in the disruptive effects induced by adolescent alcohol use
357 (Guerra and Pascual, 2019; Lamont et al., 2020; Pascual et al., 2018). Accordingly,
358 Sanchez-Marin et al. (2017) have shown that 4-week intermittent alcohol exposure (3
359 g/kg injections for 4 days/week) during adolescence induces anxiogenic-like responses
360 and impairs recognition memory later in life. Furthermore, these behavioural alterations
361 were associated with changes in the ECS and neuroinflammation-related factors.
362 Increased levels of the enzymes that mediate the synthesis of AEA and 2-AG, NAPE-
363 PLD and DAGLs, were found in the medial prefrontal cortex (mPFC) of alcohol-exposed
364 rats. In addition, an up-regulation of pro-inflammatory mediators, such as TLR4, TNF- α ,
365 COX-2 and GFAP, was induced by alcohol exposure in the mPFC. Notwithstanding,

366 lower mRNA levels of the receptors CB1 and CB2, as well as, decreased expression of
367 COX-2, GFAP, microglia response factor 1 (MRF-1) and NF- κ B were observed in the
368 striatum. Therefore, brain region-dependent changes of the ECS and neuroinflammation
369 were induced by adolescent intoxication in rats.

370 Some modulations of the eCBs have been described to confer neuroprotective effects
371 against the oxidative damage induced by binge alcohol drinking in adolescence. Selective
372 pharmacological inhibition of the FAAH enzyme, which metabolizes the N-
373 acylethanolamines, blocked the induction of oxidative stress resulting from binge alcohol
374 consumption (Pelição et al., 2016). Pre-treatment with a single dose of the FAAH
375 inhibitor URB597 (0.3 mg/kg; i.p.) was able to prevent the production of free radicals
376 after acute (3 consecutive sessions) or chronic (3 consecutive sessions over 4 weeks)
377 alcohol bingeing (6 g/kg; i.g.) in the PFC of young rats, avoiding the neurotoxicity of
378 alcohol abuse. Similarly, Bellozi et al. (2019) reported the ability of URB597 pre-
379 treatment (0.3 mg/kg; i.p.) to modulate neuroinflammation induced by binge alcohol
380 consumption during adolescence. The blockade of FAAH prevented the increase of IFN-
381 γ and TNF- α levels in the PFC and HPC induced by chronic alcohol bingeing (3 or 6 g/kg,
382 i.g.; 3 days/week for 4 weeks). Moreover, URB597 reduced the levels of IL-4, IL-10 and
383 BDNF in the PFC. Thus, the amplification of the ECS seems to exert neuroprotective
384 functions against alcohol-induced neuronal damage, albeit the exact underlying
385 mechanism needs to be further clarified. It is known that URB597 increases AEA (also
386 PEA and OEA) availability, which interacts with PPAR family receptors, known for their
387 anti-inflammatory and antioxidant activities.

388 In this context, the role of OEA, a structural analogue of AEA, has been investigated to
389 counteract the alcohol-induced damage. Using a 5-week pharmacological administration
390 of OEA (10 mg/kg/day; i.p.; 5 days/week) in a rat model of binge-like alcohol
391 consumption (3 g/kg, i.g.; weekly) combined with acute administrations of THC (5
392 mg/kg; i.p) during adolescence, Silva-Peña et al. (2019) demonstrated that OEA was
393 capable of preventing the short-term spatial memory impairments induced by alcohol and
394 THC. Furthermore, combined administration of OEA and THC restored the alcohol-
395 induced BDNF deficiency in plasma. However, OEA reduced mRNA expression of
396 BDNF in the HPC, even though it increased the expression of its receptor tropomyosin
397 receptor kinase B (TrkB). In addition, repeated administration of OEA rescued the

398 reduction of neural stem cell proliferation and newborn cell survival in the subgranular
399 zone of the dentate gyrus induced by alcohol and THC exposure and increased the
400 hippocampal levels of phospho-AKT and phospho-ERK1, key signalling regulators of
401 neurogenesis and cell survival. Also, both treatments (THC and OEA) alone or in
402 combination diminished the mRNA levels of the pro-apoptotic protease caspase-3 in the
403 HPC. Hence, OEA and other putative PPAR- α activators seem to display a protective role
404 in response to cognitive and brain dysfunctions related to alcohol exposure.

405 Recently, another approach by which memory deficits induced by adolescent alcohol
406 exposure can be rescued through the modulation of the ECS in mice has been described
407 (Peñasco et al., 2020). Repeated binge alcohol drinking (20% (v/v); 4 days/week for 4
408 weeks) during adolescence leads to long-term deficits in CB1 receptor expression and
409 distribution in the brain and consequently, it disrupts a form of excitatory long-term
410 depression that is dependent on CB1 receptors (eCB-eLTD) in the hippocampal dentate
411 gyrus. These alterations were associated with an impairment of recognition memory in
412 alcohol-exposed mice 2 weeks after the cessation of alcohol consumption, which could
413 be rescued by inhibiting the MAGL with JZL184 (8 mg/kg). Therefore, increasing the
414 availability of the endogenous 2-AG restored the functional deficits induced by
415 adolescent binge alcohol exposure.

416 However, co-exposure of WIN-55,212-2 (3 mg/kg; s.c.) with alcohol (increasing
417 concentrations of alcohol solution; 3-8% (v/v) alcohol) during early adolescence (PD 21-
418 30) increased anxiety-like behaviour (PD 35) and enhanced alcohol intake and preference
419 (6% (v/v) alcohol) in mice (up to PD 75) (Frontera et al., 2018). In addition, WIN-55,212-
420 2 treatment increased the number of dendritic ramifications in neurons of the substantia
421 nigra but lowered the number of dendritic spines in alcohol-exposed mice. These results
422 demonstrated that exposure to a cannabinoid receptor agonist during adolescence could
423 interfere with neural development affecting the neuronal morphology in key brain areas
424 that ultimately lead to functional and behavioural changes.

425 To summarize, extensive evidence has been focused on the interaction between alcohol
426 and cannabinoids along adolescence (see *Table 1*). Co-exposure of alcohol and potent
427 CB1 agonist drugs, such as THC or WIN-55,212-2, enhances alcohol-induced brain
428 damage, in addition to accentuating functional deficits regarding behavioural domains
429 like cognition. Interestingly, several studies have already reported the advantage of

430 implementing other cannabinoid-based approaches. Again, the data discussed above
431 suggests that preventing CB1 activation while acting through other targets, which are part
432 of the expanded endocannabinoid system, such as N-acylethanolamines/PPARs
433 pathways, could be a beneficial strategy to avoid alcohol-induced brain damage in this
434 developmental period.

435 **Table 1. Summary of *in vivo* preclinical evidences on the role of cannabinoid's modulation in alcohol-induced neuroinflammation, oxidative stress or**
 436 **neurodegeneration during the adolescence.**

Reference	Animal model	Alcohol administration	Cannabinoid treatment	EtOH molecular effects	Cannabinoid molecular effects	Synergic effects	Behavioural improvement/impairment
Bellozi <i>et al.</i> , (2019)	Male Wistar rats	Acute binge alcohol (3 or 6 g/kg; i.g.) for 3 consecutive days Chronic binge 3 binge cycles over 4 weeks (PD30)	URB597 0.3 mg/kg; i.p., 40 min before the alcohol administration	Chronic binge ↑ IFN- γ and TNF- α in PFC and HPC. ↑ IL-10 and BDNF in PFC.	No effects	↓ IFN- γ and TNF- α in PFC and HPC (<i>vs.</i> chronic alcohol binge group). ↓ IL-4, IL-10 and BDNF in PFC (<i>vs.</i> chronic alcohol binge group).	↑ long-term memory in novel object recognition (<i>vs.</i> acute 3 g/kg alcohol binge group).
Pelício <i>et al.</i> , (2016)	Male Wistar rats	Acute binge alcohol (3 or 6 g/kg; i.g.) for 3 consecutive days Chronic binge 3 binge cycles over 4 weeks (PD30)	URB597 0.3 mg/kg, i.p., 40 min before the alcohol administration	Acute (3 and 6 g/kg) and chronic (6 g/kg) binge ↑ production of superoxide anions (oxidative stress) in PFC.	No effects	↓ oxidative stress in PFC (<i>vs.</i> acute and chronic 6 g/kg alcohol binge).	Not applicable

Silva-Peña <i>et al.</i> , (2019)	Male Wistar rats	EtOH 3 g/kg; i.g. once per week for 5 consecutive weeks (PD34-69)	OEA 10 mg/kg; i.p.; 5 days/week, for 5 weeks Δ⁹-THC 5 mg/kg; i.p.; 1 day/week for 5 weeks Combination of both	↑ ERK1 and ERK2 in HPC ↓ pERK1/ERK1 and pERK2/ERK2 in HPC	Not applicable	OEA ↑ pAKT/AKT and pERK1/ERK1 protein levels in HPC THC + OEA ↓ Casp-3 mRNA	OEA blocks the short-term spatial-memory impairment (Y-maze) in rats induced by THC+alcohol.
Frontera <i>et al.</i> , (2018)	Male CD1 mice	Forced EtOH consumption alcohol 3% (PD21-24), alcohol 6% (PD25-27), alcohol 8% (PD 28-29).	WIN55-212,2 3mg/kg, s.c. daily administered (P30-P35)	Not applicable.	Not applicable.	↑ dendritic ramifications in neurons of the substantia nigra ↓ number of dendritic spines	↑ alcohol preference (two-bottle choice). ↑ anxiety-like behaviour in the open field.
Peñasco <i>et al.</i> , (2020)	Male C57BL/6 mice	DID test Limited access to 20% (v/v) alcohol for 4 days/week over 4 weeks (PD32-56)	JZL184 8 mg/kg, i.p. (PD 67-71)	↓ CB1 receptor-mediated excitatory transmission ↓ eCB-eLTD at medial perforant path synapses.	Not applicable.	↑ eCB-eLTD	JZL184 rescued alcohol-induced deficits in recognition memory

438 Alcohol molecular effects (alcohol group vs vehicle control group); Cannabinoid molecular effects (cannabinoid group vs vehicle control group); Synergic effect (alcohol +
439 cannabinoid group vs alcohol group). **Abbreviations:** Δ^9 -Tetrahydrocannabinol (Δ^9 -THC); Brain-derived neurotrophic factor (BDNF); Hippocampus (HPC); Interferon- γ
440 (IFN- γ); Interleukin (IL); intragastrical (i.g.); intraperitoneal (i.p.); Oleoylethanolamide (OEA); Postnatal day (PD); Prefrontal cortex (PFC); subcutaneous (s.c.); Tumour
441 necrosis factor- α (TNF- α).

442 4.2. *Adult alcohol exposure*

443 Several studies have shown that cannabinoid treatment could mitigate neuroinflammation
444 and damage produced by alcohol exposure during adulthood (see *Table 3*). In this sense,
445 OEA could prevent the neuroimmune response not only in rodent models (Antón et al.,
446 2017; Orio et al., 2019) but also in humans (Antón et al., 2018; Orio et al., 2019).
447 Preclinical studies suggest that OEA not only prevents alcohol self-administration relapse
448 and reduces withdrawal signs of alcohol (Bilbao et al., 2016) but it also inhibits the
449 expression of proinflammatory markers in brain (Antón et al., 2017). Authors applied an
450 intragastric alcohol (3g/kg) treatment to male Wistar adult rats, followed by a repeated 4-
451 day binge paradigm until they reached an intoxicating blood alcohol levels (Antón et al.,
452 2017). Consistent with this result, pre-treatment with OEA (5 mg/kg, i.p.) prevented the
453 expression of inflammatory mediators induced by alcohol exposure in the frontal cortex,
454 such as HMGB1, TLR4, the myeloid differentiation protein-2 (MD2), and MyD88.
455 Additionally, they found that OEA pre-treatment blocked the HMGB1/TLR4/MyD88
456 danger cascade associated with NF- κ B-mediated pro-inflammatory pathway and the
457 MCP-1 in frontal cortex in alcohol-binged rats (Antón et al., 2017), when the animals
458 were sacrificed 2-4h after last alcohol administration. In parallel, OEA (5mg/kg, i.p.)
459 could ameliorate alcohol-induced damage due to its neuroprotectant activity, reducing
460 caspase-3 activity, the expression of caspase 8, COX-2 and the iNOS (Antón et al., 2017),
461 molecules intimately involved with cell death and neurotoxicity associated with alcohol
462 (Liu et al., 2020; Pascual et al., 2007).

463 By contrast, other studies showed contradictory results regarding neuroinflammation
464 depending on both the dose and brain area studied. Whereas the OEA (10mg/kg, i.p.)
465 treatment *per se* increases the number of Iba1-positive cells in the HPC (Rivera et al.,
466 2018), it decreases this cell population in the striatum (Rivera et al., 2019). Additionally,
467 OEA increases the Iba1- and iNOS-positive cells but decreases the vimentin
468 immunoreactivity in the striatum of alcohol-intoxicated and control rats, indicating a
469 reduction of astrogliosis (Rivera et al., 2019). Surprisingly, their results in the striatum
470 showed a reduced reactive astroglia despite the increased number of GFAP-positive cells.
471 Authors then hypothesized that these discrepancies either could be explained due to the
472 U-shape partial and dose-dependent OEA effects or due to the OEA-induced glial
473 recruitment and/or proliferation in these specific areas (Rivera et al., 2019). When the

474 cannabinoid is co-administered with a 2 weeks of 10% alcohol liquid diet, both increase
475 the Iba1-positive cells in the striatum (Rivera et al., 2019) and HPC (Rivera et al., 2018),
476 promoting also an increasing of GFAP-positive and iNOS-positive cells in the striatum
477 (Rivera et al., 2019).

478 Moreover, OEA (5mg/kg; i.p.) is able to reduce the pro-inflammatory markers in blood.
479 Antón et al., (2017) found that this molecule could also reduce levels of TNF- α in blood
480 and IL-1 β in blood and frontal cortex of alcohol-exposed rats, after a binge alcohol
481 exposure (3g/kg; i.g.; 3 times/day for 4 days). Interestingly, a recent study in young adult
482 alcohol binge drinkers during abstinence found that these inflammatory markers (TLR4,
483 IL-1 β , COX-2, IL-6) positively correlated with the OEA levels in plasma, especially in
484 females (Antón et al., 2018). Possible discrepancies could be due to the impossibility to
485 establish which is the cause or the consequence in the relationship between inflammatory
486 markers and alcohol consumption (María Antón et al., 2018). Noteworthy, patients were
487 in abstinence and, probably, the upregulation of these biomarkers could be reflecting an
488 anti-inflammatory response.

489 In the same line of OEA, the AEA (10mg/kg; i.p.) treatment increased both astrocytes
490 iNOS-positive cells in the CA1 of HPC (Rivera et al., 2018) and GFAP- and Iba1-
491 immunoreactive cells in the striatum (Rivera et al., 2019) of alcohol-exposed rats. In fact,
492 URB597 treatment also increased the number of Iba1-positive cell in HPC and induced
493 minor morphological changes. However, the URB597 (0.3mg/kg; i.p.) treatment could
494 restore the effects of alcohol-induced inflammatory and neurodegenerative process, since
495 it was able to reduce mRNA levels of Iba1, TNF- α , IL-6 and the MCP-1 and reduce iNOS-
496 positive cells, thus improving memory function in these rats (Rivera et al., 2018). Even
497 though they found increased mRNA levels of TLR4, GFAP cells and the SDF-
498 1 α /CXCL1, they found an increasing hippocampal cell population expressing chemokine
499 receptors CX3CR1, CCR2 and CCR4. These receptors have been associated with
500 recruitment processes of immune cells. Therefore, authors hypothesized that URB597
501 (0.3mg/kg; i.p.) induced an anti-inflammatory microglial activation that might counteract
502 the alcohol-induced pro-inflammatory response (Rivera et al., 2018). These observations
503 run in parallel with the protective role of URB597 against alcohol intoxication. This
504 FAAH inhibitor might prevent the alcohol-induced neuroinflammation, since it reduces
505 alcohol consumption in the drinking in the dark paradigm, and preference after early

506 withdrawal in mice (Zhou et al., 2017). Indeed, these results are supported by an increased
507 alcohol intake and preference in mice modified with the FAAH human single-nucleotide
508 polymorphism insertion (C385A) (Zhou et al., 2016), associated with a reduction of the
509 FAAH activity and an enhancement of AEA levels in humans and mice (Dincheva et al.,
510 2015). Additionally, these effects could be blocked by a pre-treatment of a selective CB1
511 antagonist AM251 (Zhou et al., 2017, 2016). Despite its role in neuroinflammation,
512 URB597 failed to attenuate the number of necrotic cells within both dentate gyrus and
513 entorhinal cortex (Liput et al., 2017) in a rat binge model.

514 Phytocannabinoids are also important in the reduction of binge alcohol-induced brain
515 damage. Recently, Karoly et al., (2018) reported an association between circulating
516 proinflammatory cytokine IL-6 marker and alcohol consumption in alcohol regular
517 drinkers. Although variations in IL-1 β were not found due to the low severity of the
518 consumers, alcohol dependent patients have shown an association between circulating IL-
519 1 β and alcohol consumption (Leclercq et al., 2014). Extended studies revealed a different
520 pattern of cytokines expression comparing alcohol and cannabis abusing patients. Among
521 others, they observed a higher level of MCP-1, some interleukins (IL-16, IL-10, IL-309,
522 IL-12-p40, IL-15), TNF- α , tissue inhibitor of metalloproteinases 2 (TIMP-2),
523 macrophage colony-stimulating factor (MCSF) and macrophage inflammatory protein 1 α
524 (MIP-1 α) (Nair et al., 2015) obtained by array profiles from monocyte-derived dendritic
525 cells. Moreover, there is a negative association between cannabis and IL-1 β in cannabis
526 users (Karoly et al., 2018), suggesting that the phytocannabinoids contained in cannabis
527 could reduce inflammatory signals in users.

528 One of the phytocannabinoids that could have a neuroprotective action is CBD, reducing
529 apoptosis in both the entorhinal cortex and HPC of rats exposed to a binge-drinking
530 alcohol model (Hamelink et al., 2005; Liput et al., 2013). In fact, the CBD neuroprotectant
531 action observed in entorhinal cortex was similar when using a transdermal or i.p. delivery
532 (Liput et al., 2013). However, the mechanisms that could mediate CBD neuroprotective
533 effects still remain unknown due to the number of targets that CBD uses to exert its action.

534 Thus, eCBs and phytocannabinoids could promote their beneficial effects reducing
535 neuroinflammatory, neurodegenerative and apoptotic signalling. Although OEA could
536 increase some specific proinflammatory markers in a dose- and brain area-dependent
537 manner, in general, this substance seems to procamate a proliferative and/or recruitment

538 process in these areas, triggering an anti-inflammatory response. Additionally, we could
539 not discard opposite roles of cannabinoids due to different time-course in the experiments
540 or time-point in animals sacrifice. Moreover, it is necessary to consider possible different
541 sexual responses in the cannabinoid anti-inflammatory effect due to the fact that all
542 studies reported in the present section have been performed in male rats. To do so, it
543 should be mandatory performed parallel studies that asses the role of cannabinoids in
544 alcohol-induced neuroinflammatory processes in females.

545 Altogether, the results recruited above suggest that the activation of the expanded
546 endocannabinoid system could prevent alcohol-induced neurotoxic damage during
547 adulthood (see *Table 2*).

548 **Table 2. Summary of *in vivo* preclinical evidences on the role of cannabinoid's modulation in alcohol-induced neuroinflammation, oxidative stress or**
 549 **neurodegeneration at adulthood.**

Reference	Animal model	Alcohol administration	Cannabinoid treatment	Alcohol molecular effects	Cannabinoid molecular effects	Synergic effects	Behavioural improvement/impairment
Antón <i>et al.</i> (2017)	Male Wistar rats	<p>Majchrowicz binge model</p> <p>Binge alcohol (3 g/kg; i.g.) 3 times/day for 4 days.</p> <p>Control group isocaloric 5% dextrose (equivalent to 3 g/kg alcohol) 3 times/day for 4 days.</p>	<p>OEA</p> <p>5 mg/kg, i.p. previous each alcohol gavage</p>	<p>1h after alcohol administration</p> <p>↑ TNF-α, IκBα</p> <p>6h after EtOH administration</p> <p>↑p65, IκB α, COX-2</p> <p>24h after alcohol administration</p> <p>↓ TNF-α, p65, IκBα</p> <p>2-4h after alcohol administration</p> <p>↑ HMGB1, TLR4, MD2, MyD88, TNF-α, IL1b and MCP1 protein levels</p>	No effects	<p>2-4h after alcohol administration</p> <p>↓ HMGB1, TLR4, MD2, MyD88, IL1b, MCP1 and HNE protein levels</p> <p>↓ TLR4, p65, iNOS, COX-2, Caspase-8 and Caspase-3 mRNA</p> <p>↓ Caspase 3 and p65 activity</p>	OEA exerted antidepressant-like effects during acute alcohol withdrawal (forced swimming and elevated plus maze).

				<p>↑ TLR4, IκB α, iNOS, HNE, Caspase-8 and Caspase 3 mRNA</p> <p>↑ Caspase 3 activity</p>			
Liput <i>et al.</i> , (2017)	Male Sprague Dawley rats	<p>Majchrowicz binge model: alcohol 0-5 g/kg; i.g., 3 times/day for 4 days.</p> <p>Control group isocaloric dextrose; 3 times/day for 4 days.</p>	<p>URB597</p> <p>0.3 mg/kg, i.p.</p> <p>Twice daily after the third intubation of alcohol /control diet and continued for the duration of binge treatment.</p>	<p>↑ FJB+ positive cells in both DG and entorhinal cortex</p>	No effects	No effects	Not applicable
Liput <i>et al.</i> , (2013)	Male Sprague Dawley rats	<p>Majchrowicz binge model: alcohol 0-5 g/kg; i.g., 3 times/day for 4 days.</p> <p>Control group isocaloric</p>	<p>CBD</p> <p>20 mg/kg i.p. or 2.5% (w/w) transdermal gel application.</p>	<p>↑ FJB+ positive cells in entorhinal cortex</p>	Not applicable	↓ FJB+ positive cells	Not applicable

		dextrose; 3 times/day for 4 days.					
Rivera <i>et al.</i> , (2018)	Male Wistar rats	<i>Ad libitum</i> access to 11% alcohol (v/v) liquid diet for 2 weeks Control group isocaloric 14.7% (w/v) sucrose liquid diet	URB597 0.3 mg/kg; i.p. OEA 10 mg/kg; i.p. AEA 10 mg/kg; i.p. ACEA 3 mg/kg; i.p. JWH133 0.2 mg/kg; i.p.	↓ iNOS+ cells in DG, CA3 and CA1 ↑ Iba1, Tnf α , IL-6 and MCP-1 mRNA ↓ CX3CR1+ cells in the DG, CA3 and the whole HPC ↑ CCR4+ cells	All treatments: ↓ iNOS+cells within the HPC OEA, AEA, ACEA, JWH133: ↑ GFAP+cells within the HPC OEA, JWH133: ↓ Iba1+cells within the HPC URB597, AEA: ↑ Iba1+cells within the HPC URB597 ↓ GFAP+cells within the HPC ↓ Tnf α mRNA	OEA, JWH133: ↑ Iba1+cells within the HPC ACEA, JWH133: ↑ GFAP+cells in DG URB597 ↓ iNOS+cells in CA3 and CA1 and whole HPC ↓ Iba1, Tnf α , IL-6 and MCP-1 mRNA ↑ Tlr4, Gfap, Sdf-1 α /Cxcl12 mRNA ↑ CX3CR1+, CCR2+ and CCR4+ cells in the HPC ↑ CX3CR1+ cells in the DG, CA3, CA1	URB597 improves memory (novel object recognition) vs alcohol group.

					<p>↑ CX3CR1+ cells population in CA1</p> <p>↑ the CCR2+ cells population in the DG, CA3, CA1</p> <p>↑ CCR4+ cells in CA1</p> <p>↓ CXCR4+cells in DG</p>	<p>AEA</p> <p>↑ iNOS+cells in CA1</p>	
Rivera <i>et al.</i> , (2019)	Male Wistar rats	<p><i>Ad libitum</i> access to 10% alcohol (v/v) liquid diet for 2 weeks</p> <p>Control group isocaloric 14.7% (w/v) sucrose liquid diet</p>	<p>OEA 10 mg/kg, i.p.</p> <p>AEA 10 mg/kg, i.p.</p> <p>Daily treatment the last 5 days of EtOH exposure</p>	<p>In striatum:</p> <p>↑ <i>Nape-pld</i> mRNA</p> <p>↓ iNOS+ cells</p> <p>↑ Caspase- 3</p> <p>↓caspase 3 + cells expressing Iba1</p>	<p>In striatum:</p> <p>OEA</p> <p>↓ vimentin+ cells</p> <p>↑ Iba1+ and iNOS+ cells</p> <p>↓ GFAP+/cleaved caspase 3 + cells</p> <p>AEA</p> <p>↑ GFAP+ cells</p> <p>↑ Iba1+ and iNOS+ cells</p> <p>↑ Caspase-3</p>	<p>In striatum:</p> <p>OEA</p> <p>↑ GFAP+, Iba1+ and iNOS+ cells</p> <p>↑ Caspase 3</p> <p>↓ Caspase 3 + cells expressing Iba-1</p> <p>↓ GFAP+/cleaved caspase 3 + cells</p> <p>AEA</p> <p>↑ GFAP+ and Iba1+ cells</p>	<p>OEA and AEA</p> <p>↑ sucrose intake</p> <p>AEA</p> <p>↑ EtOH intake.</p> <p>OEA</p> <p>↑ alcohol -induced hypolocomotion</p>

						↓ caspase 3 + cells expressing Iba1	
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550

551 Alcohol molecular effects (alcohol group vs vehicle control group); Cannabinoid molecular effects (cannabinoid group vs vehicle control group); Synergic effect (alcohol +
552 cannabinoid group vs alcohol group). **Abbreviations:** Arachidonyl-2-chloroethylamide (ACEA); Anandamide (AEA); Cannabidiol (CBD); Cyclooxygenase-2(COX-
553 2);Chemokine receptor (CCR); CX3C chemokine receptor 1 (CX3CR1); Dentate gyrus (DG); Frontal Cortex (FC); Fluoro-Jade B (FJB); Glial fibrillary acidic protein (GFAP);
554 Glutathione (GSH); High mobility group box 1 (HMGB1); Hippocampus (HPC); Inducible nitric oxide synthase (iNOS); Ionized calcium binding adaptor molecule 1 (Iba1);
555 Inhibitory kappa B α (I κ B α); Interleukin (IL); intragastrical (i.g.); intraperitoneal (i.p.); Monocyte chemoattractant protein 1 (MCP-1); Myeloid differentiation factor (MyD88);
556 Oleoylamide (OEA); Chemokine stromal cell-derived factor 1 (SDF-1 α /CXCL1); Toll-like receptor (TLR); Tumour necrosis factor- α (TNF- α).

557 To conclude this section, it is crucial to highlight the controversial evidence regarding the
558 cannabinoid-induced harmful/therapeutic effects on the modulation of damaging
559 consequences associated with alcohol exposure. This lack of consistency among studies
560 is due to the breadth of the term “cannabinoid”, including a lot of endogenous and
561 exogenous compounds. As we mentioned before, usage of CB1 agonists commonly
562 induce detrimental outcomes, whereas non-CB1 agonist could promote desirable anti-
563 inflammatory and neuroprotective effects. Thus, further preclinical studies are needed to
564 fully clarify the mechanisms of action underlying these promising cannabinoid-based
565 drugs.

566 **5. Putative mechanisms underlying cannabinoid modulation of alcohol-induced** 567 **neuroinflammation**

568 Cannabinoids are able to modulate neuroinflammatory responses due to alcohol exposure.
569 However, the mechanisms by which cannabinoids interfere with alcohol-induced
570 intracellular signalling to counterbalance the inflammatory phenotype are complex and
571 remain poorly understood.

572 *5.1. Inhibition of pro-inflammatory signalling*

573 The cannabinoid-induced interruption of the neuroinflammatory signalling can be
574 mediated by cannabinoid receptor-dependent or -independent mechanisms as showed in
575 Figure 1A.

576 *5.1.1. Cannabinoid receptors-dependent mechanisms*

577 Despite the extensive research regarding intracellular events triggered by cannabinoid
578 receptor activation, the available information regarding putative interactions with
579 pathways regulating immune response is often contradictory and solid conclusions are
580 difficult to draw. Yang et al. (2013) found that activation of CB1 inhibits the TRPV1-
581 induced inflammatory response in corneal epithelial cells. Upon CB1 agonism, inhibition
582 of cyclic adenosine monophosphate (cAMP) formation decreases the cAMP-dependent
583 activation of protein kinase A (PKA). This decline translates in a reduction of the TRPV1
584 phosphorylation and activation, leading to a blunting in the inflammatory response.
585 Ehrhart et al. (2005) demonstrated that the agonism of CB2 inhibits the IFN- γ -induced

586 phosphorylation of JAK/STAT1 and decreases microglial production of proinflammatory
587 mediators, such as TNF- α and nitric oxide. JAK/STAT signalling is the main molecular
588 pathway activated by ILs (Murray, 2007), promoting the expression of either pro-
589 inflammatory or anti-inflammatory genes. Noteworthy, the most documented STAT upon
590 the inflammatory signalling are the subtype STAT1 and STAT3. STAT1 induces the
591 expression of pro-inflammatory molecules, whereas STAT3 mostly activates the
592 expression of suppressors of cytokine signalling (SOCS) (Carey et al., 2012; Murray,
593 2006). However, the role of STAT3 in inflammation is still controversial, as the activation
594 of this transcriptional factor has been shown to both repress the transcription of pro-
595 inflammatory genes and promote cell apoptosis (Nabavi et al., 2019). Another study
596 revealed that AEA-dependent microglial CB2 activation inhibits the phosphorylation of
597 inhibitory kappa B α (I κ B α), preventing the translocation of NF- κ B to the nucleus (Correa
598 et al., 2010). In the same line, the activation of cannabinoid receptors, specially CB2, is
599 related to the drive of activated microglia to more homeostatic phenotypes (Mecha et al.,
600 2016). Therefore, cannabinoid receptors, through the inhibition of different components
601 of the inflammatory signalling, might ameliorate the immune response caused by alcohol
602 exposure.

603 *5.1.2. Cannabinoid receptors-independent mechanisms*

604 PPAR α and PPAR γ are involved in the inhibition of the NF- κ B and AP-1 pro-
605 inflammatory pathways, which are highly involved in the alcohol-induced inflammatory
606 response. When activated, these nuclear receptors are able to translocate to the nucleus
607 and bind to transcription factors to prevent their activity through a mechanism called
608 transrepression. Another form of transrepression is the binding of PPARs to the repressor
609 complex located on the promoter of inflammatory genes, therefore preventing their
610 transcription by NF- κ B or AP-1 (Varga et al., 2011b). Du et al. (2011) inhibited the
611 phosphorylation of NF- κ B and further COX-2 expression through the activation of
612 PPAR γ by 2-AG and other PPAR γ synthetic agonists in hippocampal neurons *in vitro*.
613 Another study by Yang et al. (2016) revealed that the activation of PPAR α by OEA
614 inhibits phosphorylation of I κ B α , preventing the activation of NF- κ B, in THP-1
615 monocytic cells. In the same study, activation of PPAR α by OEA was also found to inhibit
616 the ERK1/2/AP-1/STAT3 pathway, leading to the suppression of the inflammatory
617 response.

618 On the other side of the balance, TRPV1 stimulation leads to the activation of pro-
619 inflammatory signalling pathways and a potential increase of alcohol-induce
620 neuroinflammation. Therefore, the modulation of this receptor could be of therapeutic
621 interest. When activated, intracellular Ca²⁺ rises in a TRPV-1 dependent manner leading
622 to the phosphorylation of ERK1/2 and the activation of the AP-1 transcription factor
623 (Backes et al., 2018) in H2C1 cell line. Furthermore, the TRPV1 stimulation-induced
624 increase in intracellular Ca²⁺ is thought to activate the transforming growth factor beta-
625 activated kinase 1 (TAK1) in human corneal epithelial cells, which will ultimately lead
626 to the activation of NF-κB and AP-1 (Wang et al., 2011; Yang et al., 2013), although this
627 mechanism needs to be further explored. Nevertheless, some studies have found
628 activation of TRPV1 to play neuroprotective and anti-inflammatory roles, although the
629 mechanism of action are not clear (Kong et al., 2017).

630 Microglial GPR55 stimulation has also been shown to activate ERK1/2 and NF-κB,
631 leading to the transcription of pro-inflammatory mediators (Liu et al., 2015; Pietr et al.,
632 2009). Therefore, the antagonism of GPR55 by cannabinoids, such as CBD (Kaplan et
633 al., 2017), could lead to more protective and homeostatic phenotypes to counterbalance
634 the alcohol-induced neuroinflammation.

635 In the CNS, alcohol-induced release of HMGB1 and other DAMPs activates TLR4, which
636 triggers different intracellular pathways leading to the activation of the main pro-
637 inflammatory transcription factors, NF-κB, AP-1 and IRF3, as well as STAT1. As we
638 have reviewed in this section, cannabinoids are able to interfere with the proper course of
639 this signalling in order to inhibit the pro-inflammatory response.

640 *5.2. Activation of anti-inflammatory signalling*

641 Despite a broad body of research that has focused on the pro-inflammatory signalling
642 inhibition, molecular mechanisms that regulate the transition of microglia from
643 detrimental states to phenotypes associated with CNS homeostasis are currently being
644 investigated. Modulation of the ECS has been proposed as a promising strategy against
645 pro-inflammatory microglial activation (Cristino et al., 2020; Franco and Fernández-
646 Suárez, 2015), as represented in Figure 1B.

647 *5.2.1. Upregulation of anti-inflammatory molecules*

648 Evidence reveals that many cannabinoids can induce the expression of anti-inflammatory
649 cytokines, such as IL-4, IL-10 or TFG- β , in a chronic inflammatory context (Al-Ghezi et
650 al., 2019; Smith et al., 2000), which might be partially mediated by CB2 (Correa et al.,
651 2010; Robinson et al., 2015). Although depending on different factors like the
652 inflammatory context, generally anti-inflammatory ILs bring balance toward STAT3 or
653 STAT6 activation, enhancing the function of suppressors of cytokine signalling (SOCS)
654 (Busch-Dienstfertig and González-Rodríguez, 2013; Carey et al., 2012). SOCS
655 expression mainly leads to the inhibition of the cytokine-JAK/STAT signalling, as a
656 negative feedback mechanism (Liau et al., 2018). In this respect, one study displayed that
657 CBD was able to strengthen the STAT3 activation in a LPS-induced pro-inflammatory
658 response, thereby promoting anti-inflammatory signalling in BV-2 microglia cells
659 (Kozela et al., 2010).

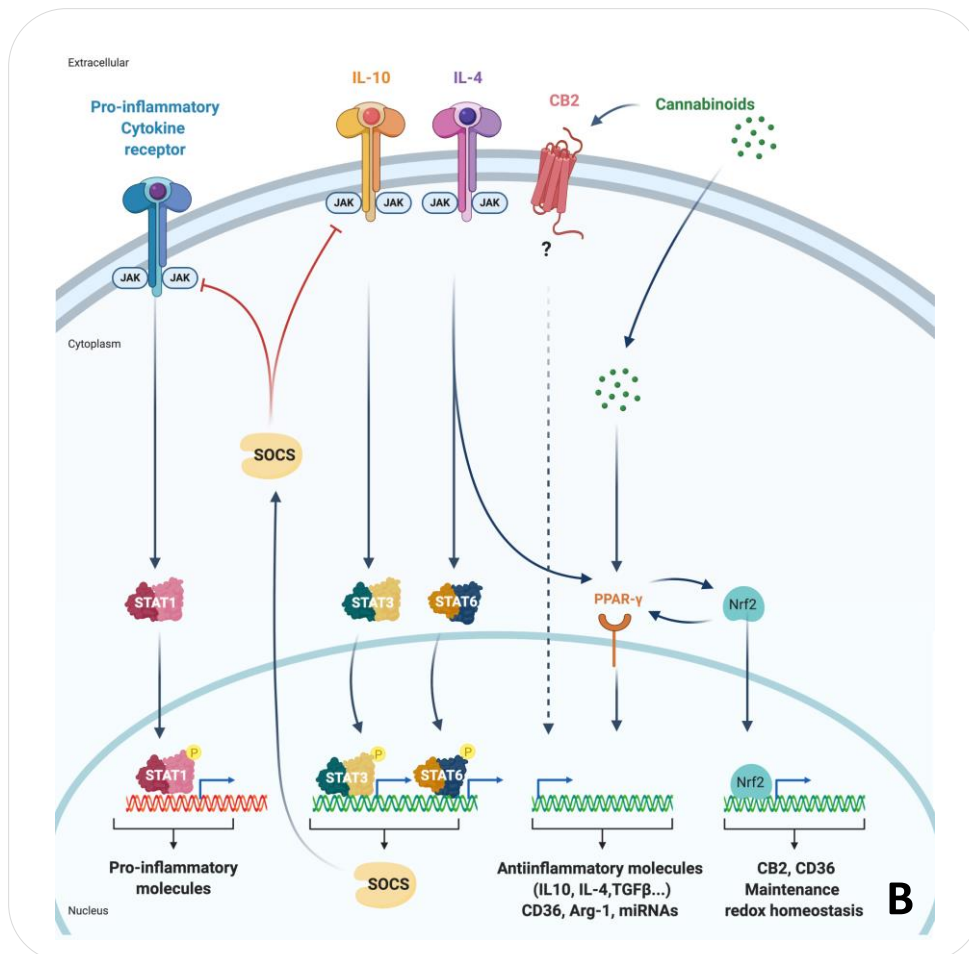
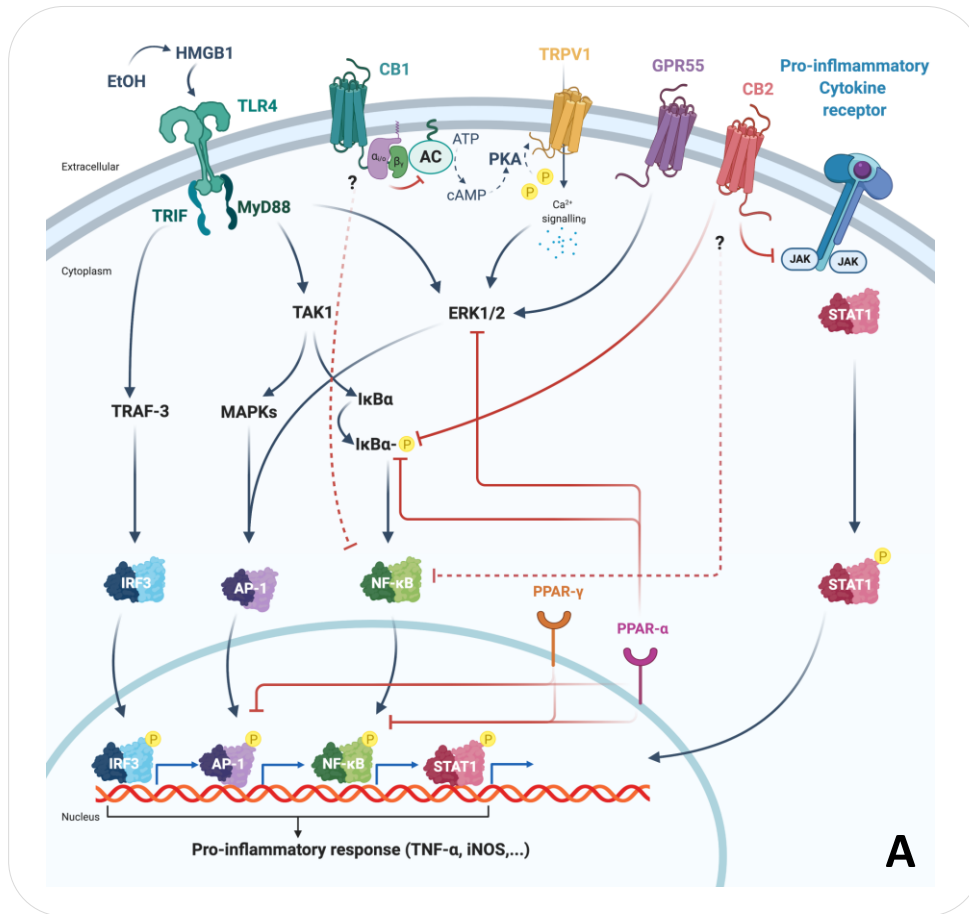
660 In addition, other authors have suggested non-canonical pathways by which the anti-
661 inflammatory IL-4 could directly activate the PPAR γ (Li et al., 2014), which then
662 translocates to the nucleus to accomplish its biological function. In turn, some studies
663 revealed that the activation of these nuclear receptors by a specific agonist enhanced some
664 anti-inflammatory IL, such as IL-10, in microglia (Choi et al., 2017a, 2017b).
665 Nonetheless, other molecular pathways in which PPAR γ is a key modulator could be
666 exploited.

667 *5.2.2. Activation of PPAR γ*

668 The PPAR γ activation increases the expression of Arginase-1, which is a pro-
669 regenerative microglial marker (Fumagalli et al., 2018). Besides, a positive loop exists
670 between PPAR γ and nuclear factor erythroid 2-related factor 2 (Nrf2). Therefore, PPAR γ
671 can upregulate Nrf2 and vice versa. Nrf2 is a transcriptional factor that activates the
672 expression of numerous genes critical for the maintenance of redox homeostasis (Zhang
673 et al., 2013). These proteins are necessary to inhibit the generation of reactive oxygen
674 species (ROS) and mitigate neuroinflammation (Cai et al., 2018; Rojo et al., 2018).
675 Moreover, CD36 microglial marker, which is under the regulation of both PPAR γ and
676 Nrf2 (Yamanaka et al., 2012; Zhao et al., 2015), has been shown to facilitate the
677 resolution of oxidative stress and neuroinflammation (Ballesteros et al., 2014; Huang et
678 al., 2014). Interestingly, a recent study has demonstrated that the expression of CB2 in

679 microglia is Nrf2-dependent (Galán-Ganga et al., 2020). As we mentioned above, CB2
680 agonism has also been proved to enhance anti-inflammatory IL expression (Correa et al.,
681 2010; Robinson et al., 2015). In this context, it is established that both the exogenous
682 cannabinoids and the expanded endocannabinoid molecules can activate the PPAR γ
683 (O'Sullivan, 2016) as well as the CB2. Furthermore, the modulation of these two
684 receptors has been proposed to exert neuroprotective and anti-inflammatory effects in the
685 alcohol context (Cippitelli et al., 2017; Nair et al., 2015). Thus, distinct cannabinoids
686 might be underpinning a promising mechanism involving PPAR γ -Nrf2-CB2 and anti-
687 inflammatory cytokines, to switch to alternative and pro-regenerative microglial
688 activation.

689 In the last decades, an epigenetic mechanism which involved the link between
690 microRNAs (miRNA) expression and alcohol-related disorders has been explored
691 (Ignacio et al., 2015; Mandal et al., 2018). Some studies have suggested that the
692 modulation of miRNAs expression plays a key role in the pathophysiologic inflammation
693 induced by alcohol exposure (Lippai et al., 2014, 2013). In regard to the inflammatory
694 response, other authors have proposed the implication of several miRNAs in promoting
695 pro-regenerative microglia phenotypes (Fumagalli et al., 2018; Guo et al., 2019). In fact,
696 a mechanistic study has confirmed that inhibition of pro-inflammatory cytokines
697 expression by the upregulation of miRNA-124 is mediated by the activation of PPAR γ
698 (Wang et al., 2017). More evidence reported the effect of distinct miRNAs on the
699 microglia polarization, highlighting that miRNA-124 could promote the anti-
700 inflammatory phenotype (Guo et al., 2019). Interestingly, a recently reported mechanism
701 shows that cannabinoids can interfere with neuroinflammation through the modulation of
702 several miRNAs (Dinu et al., 2020). Another research group has identified a repertoire of
703 miRNAs that are regulated by cannabinoids in resting and LPS-activated microglia
704 (Juknat et al., 2019). Their results revealed that the modulated miRNAs are linked to
705 inflammatory pathways and Nrf2-mediated cellular stress.



707 **Figure 1. Cannabinoid modulation of alcohol-induced neuroinflammation in microglia. A) Inhibition**
708 **of pro-inflammatory signalling.** Cannabinoid binding to different receptors from the expanded
709 endocannabinoid system is able to interrupt pro-inflammatory signalling pathways. CB1 and CB2
710 activation leads to the inhibition of NF- κ B. Besides, CB1 inhibits the TRPV1 and CB2 inhibits the
711 JAK/STAT1 pro-inflammatory signalling pathways. PPAR α and PPAR γ directly inhibit NF- κ B and AP-1.
712 PPAR α also blocks the phosphorylation of ERK1/2. **B) Anti-inflammatory signalling activated by**
713 **cannabinoids.** Several endocannabinoids and exogenous cannabinoids can increase the expression of anti-
714 inflammatory cytokines (IL-10, IL-4, etc.) mainly through the activation of CB2 and PPAR γ . These
715 cytokines activate JAK/STAT toward an anti-inflammatory signalling, inducing the inhibition of pro-
716 inflammatory cytokines owing to the rising SOCS expression. PPAR γ is able to activate Nrf2 and vice
717 versa, promoting a positive anti-inflammatory feedback that allows an increasing expression of pro-
718 regenerative mediators (CD36, Arg-1, redox-related enzymes, miRNAs, among some anti-inflammatory
719 cytokines). Abbreviations: activator protein 1 (AP-1); arginase-1 (Arg-1); cannabinoid receptor 1 (CB1);
720 cannabinoid receptor 2 (CB2); extracellular signal-regulated kinase (ERK); interleukin (IL); interferon
721 regulatory factor 3 (IRF3); Janus kinase (JAK); nuclear factor erythroid 2-related factor 2 (Nrf2); nuclear
722 factor kappa B (NF- κ B); microRNA (miRNA); peroxisome proliferator-activated receptor (PPAR);
723 suppressor cytokine signalling (SOCS); signal transducer and activator transcription proteins (STAT);
724 transforming growth factor beta (TFG β); transient receptor potential vanilloid 1 (TRPV1). Figure created
725 with Biorender.com

726 In this section, we have hypothesised diverse mechanisms by which cannabinoid
727 modulation may exert its anti-inflammatory effects in an alcohol context. On the one
728 hand, we focused our research on the convergent molecules between pro-inflammatory
729 pathways activated by alcohol in which cannabinoids might interfere, such as NF- κ B. On
730 the other hand, we discussed complementary mechanisms based on the activation of anti-
731 inflammatory signalling which might cooperate to resolve the chronic deleterious
732 neuroinflammation caused by alcohol exposure. The studies mentioned above indicate
733 that cannabinoids are generally able to inhibit several components of pro-inflammatory
734 pathways that are activated after different insults. Furthermore, a number of studies have
735 shown the transition from detrimental microglial activation toward a regenerative
736 phenotype instigated by cannabinoid modulation. Yet, the majority of these studies have
737 been carried out upon common inflammatory insults and the factor that induces the
738 neuroinflammation is noteworthy for the type of pro-inflammatory pathways, which will
739 be activated. In this sense, further research is required to clarify the role of cannabinoids
740 in the field of alcohol-induced neuroinflammation, because there is scarce scientific
741 literature concerning mechanistic studies of how cannabinoids might act within alcohol-
742 dependent pro-inflammatory context.

743 **6. Conclusions**

744 Alcohol exposure might trigger a series of pro-inflammatory and neurodegenerative
745 signalling pathways that drives the CNS to chronic and non-physiological stages. The
746 ECS is known to play a crucial role in the modulation of neuroinflammatory processes,
747 trending towards the resolution of the immune response. Although many studies have
748 focused on the anti-inflammatory properties of cannabinoids, only a few have used
749 alcohol as the etiological factor. Despite the fact that alcohol exposure derived effects
750 vary depending on the developmental state, we have found some consistencies across
751 adolescent and adult exposures to alcohol. Evidence suggests that direct agonism to CB1
752 is related to a worsening of the alcohol-induced detrimental effects. However, other
753 therapeutic approaches based on other components of the expanded endocannabinoid
754 system, such as PPAR γ or the usage of OEA, have been shown to exert more promising
755 outcomes. The mechanisms through which cannabinoids are able to ameliorate the
756 alcohol-induced neuroinflammation are poorly understood. In this review, we propose a
757 model where cannabinoids might counterbalance the immune response in alcohol
758 contexts through the inhibition of pro-inflammatory and the stimulation of anti-
759 inflammatory signalling pathways. Therefore, cannabinoids act through multiple and
760 complementary mechanisms in order to achieve a wider and more potent effect.
761 Cannabinoid modulation represents an extremely interesting therapeutic target in alcohol-
762 induced chronic neuroinflammation.

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775 **7. References**

- 776 Achur, R.N., Freeman, W.M., Vrana, K.E., 2010. Circulating cytokines as biomarkers
777 of alcohol abuse and alcoholism. *J. Neuroimmune Pharmacol.* 5, 83-91.
778 <https://doi.org/10.1007/s11481-009-9185-z>
- 779 Agudelo, M., Yndart, A., Morrison, M., Figueroa, G., Muñoz, K., Samikkannu, T., Nair,
780 M.P., 2013. Differential expression and functional role of cannabinoid genes in
781 alcohol users. *Drug Alcohol Depend.* 133, 789–793.
782 <https://doi.org/10.1016/j.drugalcdep.2013.08.023>
- 783 Al-Ghezi, Z.Z., Miranda, K., Nagarkatti, M., Nagarkatti, P.S., 2019. Combination of
784 cannabinoids, Δ^9 -tetrahydrocannabinol and cannabidiol, ameliorates experimental
785 multiple sclerosis by suppressing neuroinflammation through regulation of
786 miRNA-mediated signaling pathways. *Front. Immunol.* 10, 1921.
787 <https://doi.org/10.3389/fimmu.2019.01921>
- 788 Alen, F., Decara, J., Brunori, G., You, Z.-B., Bühler, K.-M., López-Moreno, J.A.,
789 Cippitelli, A., Pavon, F.J., Suárez, J., Gardner, E.L., de la Torre, R., Ciccocioppo,
790 R., Serrano, A., Rodríguez de Fonseca, F., 2018. PPAR α /CB1 receptor dual
791 ligands as a novel therapy for alcohol use disorder: Evaluation of a novel oleic acid
792 conjugate in preclinical rat models. *Biochem. Pharmacol.* 157, 235–243.
793 <https://doi.org/10.1016/j.bcp.2018.09.008>
- 794 Alfonso-Loeches, S., Pascual-Lucas, M., Blanco, A.M., Sanchez-Vera, I., Guerri, C.,
795 2010. Pivotal Role of TLR4 Receptors in Alcohol-Induced Neuroinflammation and
796 Brain Damage. *J. Neurosci.* 30, 8285–8295.
797 <https://doi.org/10.1523/JNEUROSCI.0976-10.2010>
- 798 Alfonso-Loeches, S., Pascual, M., Guerri, C., 2013. Gender differences in alcohol-
799 induced neurotoxicity and brain damage. *Toxicology* 311, 27–34.
800 <https://doi.org/10.1016/j.tox.2013.03.001>
- 801 Alfonso-Loeches, S., Ureña-Peralta, J., Morillo-Bargues, M.J., Gómez-Pinedo, U.,
802 Guerri, C., 2016. Ethanol-Induced TLR4/NLRP3 Neuroinflammatory Response in
803 Microglial Cells Promotes Leukocyte Infiltration Across the BBB. *Neurochem.*

- 804 Res. 41, 193–209. <https://doi.org/10.1007/s11064-015-1760-5>
- 805 Antón, M., Alén, F., Gómez de Heras, R., Serrano, A., Pavón, F.J., Leza, J.C., García-
806 Bueno, B., Rodríguez de Fonseca, F., Orió, L., 2017. Oleoylethanolamide prevents
807 neuroimmune HMGB1/TLR4/NF-kB danger signaling in rat frontal cortex and
808 depressive-like behavior induced by ethanol binge administration. *Addict. Biol.* 22,
809 724–741. <https://doi.org/10.1111/adb.12365>
- 810 Antón, M, Rodríguez-González, A., Ballesta, A., González, N., del Pozo, A., de
811 Fonseca, F.R., Gómez-Lus, M.L., Leza, J.C., García-Bueno, B., Caso, J.R., Orió,
812 L., 2018. Alcohol binge disrupts the rat intestinal barrier: the partial protective role
813 of oleoylethanolamide. *Br. J. Pharmacol.* 175, 4464–4479.
814 <https://doi.org/10.1111/bph.14501>
- 815 Antón, M., Rodríguez-González, A., Rodríguez-Rojo, I.C., Pastor, A., Correas, Á.,
816 Serrano, A., Ballesta, A., Alén, F., Gómez de Heras, R., de la Torre, R., Rodríguez
817 de Fonseca, F., Orió, L., 2018. Increased plasma oleoylethanolamide and
818 palmitoleoylethanolamide levels correlate with inflammatory changes in alcohol
819 binge drinkers: the case of HMGB1 in women. *Addict. Biol.* 23, 1242–1250.
820 <https://doi.org/10.1111/adb.12580>
- 821 Aoki, T., Narumiya, S., 2012. Prostaglandins and chronic inflammation. *Trends*
822 *Pharmacol. Sci.* <https://doi.org/10.1016/j.tips.2012.02.004>
- 823 Araujo, D.J., Tjoa, K., Saijo, K., 2019. The Endocannabinoid System as a Window Into
824 Microglial Biology and Its Relationship to Autism. *Front. Cell. Neurosci.*
825 <https://doi.org/10.3389/fncel.2019.00424>
- 826 Atwood, B.K., Mackie, K., 2010. CB2: a cannabinoid receptor with an identity crisis.
827 *Br. J. Pharmacol.* 160, 467–479. <https://doi.org/10.1111/j.1476-5381.2010.00729.x>
- 828 Aymerich, M.S., Aso, E., Abellanas, M.A., Tolon, R.M., Ramos, J.A., Ferrer, I.,
829 Romero, J., Fernández-Ruiz, J., 2018. Cannabinoid pharmacology/therapeutics in
830 chronic degenerative disorders affecting the central nervous system. *Biochem.*
831 *Pharmacol.* 157, 67-84. <https://doi.org/10.1016/j.bcp.2018.08.016>
- 832 Backes, T.M., Rössler, O.G., Hui, X., Grötzinger, C., Lipp, P., Thiel, G., 2018.

833 Stimulation of TRPV1 channels activates the AP-1 transcription factor. *Biochem.*
834 *Pharmacol.* 150, 160–169. <https://doi.org/10.1016/j.bcp.2018.02.008>

835 Ballesteros, I., Cuartero, M.I., Pradillo, J.M., de la Parra, J., Perez-Ruiz, A., Corbi, A.,
836 Ricote, M., Hamilton, J.A., Sobrado, M., Vivancos, J., Nombela, F., Lizasoain, I.,
837 Moro, M.A., 2014. Rosiglitazone-induced CD36 up-regulation resolves
838 inflammation by PPAR and 5-LO-dependent pathways. *J. Leukoc. Biol.* 95, 587–
839 598. <https://doi.org/10.1189/jlb.0613326>

840 Banks, W.A., Erickson, M.A., 2010. The blood-brain barrier and immune function and
841 dysfunction. *Neurobiol. Dis.* <https://doi.org/10.1016/j.nbd.2009.07.031>

842 Bassi, M.S., Gentile, A., Iezzi, E., Zagaglia, S., Musella, A., Simonelli, I., Gilio, L.,
843 Furlan, R., Finardi, A., Marfia, G.A., Guadalupi, L., Bullitta, S., Mandolesi, G.,
844 Centonze, D., Buttari, F., 2019. Transient receptor potential vanilloid 1 modulates
845 central inflammation in multiple sclerosis. *Front. Neurol.* 10.
846 <https://doi.org/10.3389/fneur.2019.00030>

847 Bátkai, S., Mukhopadhyay, P., Horváth, B., Rajesh, M., Gao, R.Y., Mahadevan, A.,
848 Amere, M., Battista, N., Lichtman, A.H., Gauson, L.A., Maccarrone, M., Pertwee,
849 R.G., Pacher, P., 2012. Δ^8 -Tetrahydrocannabivarin prevents hepatic
850 ischaemia/reperfusion injury by decreasing oxidative stress and inflammatory
851 responses through cannabinoid CB2 receptors. *Br. J. Pharmacol.* 165, 2450–2461.
852 <https://doi.org/10.1111/j.1476-5381.2011.01410.x>

853 Bellozi, P.M.Q., Pelicão, R., Santos, M.C., Lima, I.V.A., Saliba, S.W., Vieira, É.L.M.,
854 Campos, A.C., Teixeira, A.L., de Oliveira, A.C.P., Nakamura-Palacios, E.M.,
855 Rodrigues, L.C.M., 2019. URB597 ameliorates the deleterious effects induced by
856 binge alcohol consumption in adolescent rats. *Neurosci. Lett.* 711, 134408.
857 <https://doi.org/10.1016/j.neulet.2019.134408>

858 Bilbao, A., Serrano, A., Cippitelli, A., Pavón, F.J., Giuffrida, A., Suárez, J., García-
859 Marchena, N., Baixeras, E., Gómez de Heras, R., Orio, L., Alén, F., Ciccocioppo,
860 R., Cravatt, B.F., Parsons, L.H., Piomelli, D., Rodríguez de Fonseca, F., 2016.
861 Role of the satiety factor oleoylethanolamide in alcoholism. *Addict. Biol.* 21, 859–
862 872. <https://doi.org/10.1111/adb.12276>

- 863 Bisogno, T., Howell, F., Williams, G., Minassi, A., Cascio, M.G., Ligresti, A., Matias,
864 I., Schiano-Moriello, A., Paul, P., Williams, E.-J., Gangadharan, U., Hobbs, C., Di
865 Marzo, V., Doherty, P., 2003. Cloning of the first sn1-DAG lipases points to the
866 spatial and temporal regulation of endocannabinoid signaling in the brain. *J. Cell*
867 *Biol.* 163, 463–8. <https://doi.org/10.1083/jcb.200305129>
- 868 Blednov, Y.A., Benavidez, J.M., Black, M., Ferguson, L.B., Schoenhard, G.L., Goate,
869 A.M., Edenberg, H.J., Wetherill, L., Hesselbrock, V., Foroud, T., Adron Harris, R.,
870 2015. Peroxisome Proliferator-Activated Receptors α and γ are Linked with
871 Alcohol Consumption in Mice and Withdrawal and Dependence in Humans.
872 *Alcohol. Clin. Exp. Res.* 39, 136–145. <https://doi.org/10.1111/acer.12610>
- 873 Busch-Dienstfertig, M., González-Rodríguez, S., 2013. IL-4, JAK-STAT signaling, and
874 pain. *JAK-STAT* 2, e27638. <https://doi.org/10.4161/jkst.27638>
- 875 Cabral, G.A., Griffin-Thomas, L., 2009. Emerging role of the cannabinoid receptor CB
876 2 in immune regulation: therapeutic prospects for neuroinflammation. *Expert Rev.*
877 *Mol. Med.* 11, e3. <https://doi.org/10.1017/S1462399409000957>
- 878 Cai, W., Yang, T., Liu, H., Han, L., Zhang, K., Hu, X., Zhang, X., Yin, K.-J., Gao, Y.,
879 Bennett, M.V.L., Leak, R.K., Chen, J., 2018. Peroxisome proliferator-activated
880 receptor γ (PPAR γ): A master gatekeeper in CNS injury and repair. *Prog.*
881 *Neurobiol.* 163–164, 27–58. <https://doi.org/10.1016/j.pneurobio.2017.10.002>
- 882 Capuron, L., Miller, A.H., 2011. Immune system to brain signaling:
883 Neuropsychopharmacological implications. *Pharmacol. Ther.* 226-38.
884 <https://doi.org/10.1016/j.pharmthera.2011.01.014>
- 885 Carey, A.J., Tan, C.K., Ulett, G.C., 2012. Infection-induced IL-10 and JAK-STAT.
886 *JAK-STAT* 1, 159–167. <https://doi.org/10.4161/jkst.19918>
- 887 Cassano, T., Calcagnini, S., Pace, L., Marco, F. De, Romano, A., Gaetani, S., 2017.
888 Cannabinoid receptor 2 signaling in neurodegenerative disorders: From
889 pathogenesis to a promising therapeutic target. *Front. Neurosci.*
890 <https://doi.org/10.3389/fnins.2017.00030>
- 891 Chiurchiù, V., Leuti, A., Maccarrone, M., 2018. Bioactive lipids and chronic

- 892 inflammation: Managing the fire within. *Front. Immunol.*
893 <https://doi.org/10.3389/fimmu.2018.00038>
- 894 Chiurchiù, V., Leuti, A., Maccarrone, M., 2015. Cannabinoid Signaling and
895 Neuroinflammatory Diseases: A Melting pot for the Regulation of Brain Immune
896 Responses. *J. Neuroimmune Pharmacol.* 10, 268–280.
897 <https://doi.org/10.1007/s11481-015-9584-2>
- 898 Choi, M.J., Lee, E.J., Park, J.S., Kim, S.N., Park, E.M., Kim, H.S., 2017a. Anti-
899 inflammatory mechanism of galangin in lipopolysaccharide-stimulated microglia:
900 Critical role of PPAR- γ signaling pathway. *Biochem. Pharmacol.* 144, 120–131.
901 <https://doi.org/10.1016/j.bcp.2017.07.021>
- 902 Choi, M.J., Park, J.S., Park, J.E., Kim, Han Su, Kim, Hee Sun, 2017b. Galangin
903 suppresses pro-inflammatory gene expression in polyinosinic-polycytidylic acid-
904 stimulated microglial cells. *Biomol. Ther.* 25, 641–647.
905 <https://doi.org/10.4062/biomolther.2017.173>
- 906 Cippitelli, A., Domi, E., Ubaldi, M., Douglas, J.C., Li, H.W., Demopoulos, G.,
907 Gaitanaris, G., Roberto, M., Drew, P.D., Kane, C.J.M., Ciccocioppo, R., 2017.
908 Protection against alcohol-induced neuronal and cognitive damage by the PPAR γ
909 receptor agonist pioglitazone. *Brain. Behav. Immun.* 64, 320–329.
910 <https://doi.org/10.1016/j.bbi.2017.02.001>
- 911 Coleman, L.G., Crews, F.T., 2018. Innate Immune Signaling and Alcohol Use
912 Disorders. *Handbook of Experimental Pharmacology.* Springer New York LLC.
913 369–396. https://doi.org/10.1007/164_2018_92
- 914 Correa, F., Docagne, F., Mestre, L., Loría, F., Hernangómez, M., Borrell, J., Guaza, C.,
915 2007. Cannabinoid system and neuroinflammation: Implications for multiple
916 sclerosis. *NeuroImmunoModulation.* 182–187. <https://doi.org/10.1159/000110644>
- 917 Correa, F., Hernangómez, M., Mestre, L., Loría, F., Spagnolo, A., Docagne, F., Di
918 Marzo, V., Guaza, C., 2010. Anandamide enhances IL-10 production in activated
919 microglia by targeting CB 2 receptors: Roles of ERK1/2, JNK, and NF- κ B. *Glia*
920 58, 135–147. <https://doi.org/10.1002/glia.20907>

- 921 Cravatt, B.F., Giang, D.K., Mayfield, S.P., Boger, D.L., Lerner, R.A., Gilula, N.B.,
922 1996. Molecular characterization of an enzyme that degrades neuromodulatory
923 fatty-acid amides. *Nature* 384, 83–87. <https://doi.org/10.1038/384083a0>
- 924 Crews, F.T., Bechara, R., Brown, L.A., Guidot, D.M., Mandrekar, P., Oak, S., Qin, L.,
925 Szabo, G., Wheeler, M., Zou, J., 2006. Cytokines and alcohol. *Alcoholism:*
926 *Clinical and Experimental Research*. 30, 720–730. [https://doi.org/10.1111/j.1530-](https://doi.org/10.1111/j.1530-0277.2006.00084.x)
927 [0277.2006.00084.x](https://doi.org/10.1111/j.1530-0277.2006.00084.x)
- 928 Crews, F.T., Vetreno, R.P., 2016. Mechanisms of neuroimmune gene induction in
929 alcoholism. *Psychopharmacology (Berl)*. 233, 1543–57.
930 <https://doi.org/10.1007/s00213-015-3906-1>
- 931 Crews, F.T., Vetreno, R.P., 2014. Neuroimmune basis of alcoholic brain damage. *Int.*
932 *Rev. Neurobiol.* 118, 315–57. [https://doi.org/10.1016/B978-0-12-801284-0.00010-](https://doi.org/10.1016/B978-0-12-801284-0.00010-5)
933 [5](https://doi.org/10.1016/B978-0-12-801284-0.00010-5)
- 934 Cristino, L., Bisogno, T., Di Marzo, V., 2020. Cannabinoids and the expanded
935 endocannabinoid system in neurological disorders. *Nat. Rev. Neurol.* 16, 9–29.
936 <https://doi.org/10.1038/s41582-019-0284-z>
- 937 Croxford, J.L., Yamamura, T., 2005. Cannabinoids and the immune system: Potential
938 for the treatment of inflammatory diseases? *J. Neuroimmunol.* 166, 3–18.
939 <https://doi.org/10.1016/j.jneuroim.2005.04.023>
- 940 Cruz, C., Meireles, M., Silva, S.M., 2017. Chronic ethanol intake induces partial
941 microglial activation that is not reversed by long-term ethanol withdrawal in the rat
942 hippocampal formation. *Neurotoxicology* 60, 107–115.
943 <https://doi.org/10.1016/J.NEURO.2017.04.005>
- 944 D’Mello, C., Swain, M.G., 2017. Immune-to-brain communication pathways in
945 inflammation-associated sickness and depression. *Current Topics in Behavioral*
946 *Neurosciences*. Springer Verlag. 31, 73–94. https://doi.org/10.1007/7854_2016_37
- 947 Dantzer, R., O’Connor, J.C., Freund, G.G., Johnson, R.W., Kelley, K.W., 2008. From
948 inflammation to sickness and depression: When the immune system subjugates the
949 brain. *Nat. Rev. Neurosci.* 9, 46-56. <https://doi.org/10.1038/nrn2297>

950 De Petrocellis, L., Ligresti, A., Moriello, A.S., Allarà, M., Bisogno, T., Petrosino, S.,
951 Stott, C.G., Di Marzo, V., 2011. Effects of cannabinoids and cannabinoid-enriched
952 Cannabis extracts on TRP channels and endocannabinoid metabolic enzymes. *Br.*
953 *J. Pharmacol.* 163, 1479–94. <https://doi.org/10.1111/j.1476-5381.2010.01166.x>

954 de Timary, P., Stärkel, P., Delzenne, N.M., Leclercq, S., 2017. A role for the peripheral
955 immune system in the development of alcohol use disorders? *Neuropharmacology*
956 122, 148–160. <https://doi.org/10.1016/j.neuropharm.2017.04.013>

957 Dennis, E.A., Norris, P.C., 2015. Eicosanoid storm in infection and inflammation. *Nat.*
958 *Rev. Immunol.* 15, 511-523. <https://doi.org/10.1038/nri3859>

959 Devane, W.A., Dysarz, F.A., Johnson, M.R., Melvin, L.S., Howlett, A.C., 1988.
960 Determination and characterization of a cannabinoid receptor in rat brain. *Mol.*
961 *Pharmacol.* 34, 605–613.

962 Di Marzo, V., 2018. New approaches and challenges to targeting the endocannabinoid
963 system. *Nat. Rev. Drug Discov.* 17, 623-639. <https://doi.org/10.1038/nrd.2018.115>

964 Di Marzo, V., Bisogno, T., De Petrocellis, L., 2000. Endocannabinoids: new targets for
965 drug development. *Curr. Pharm. Des.* 6, 1361–1380.

966 Di Marzo, V., Melck, D., Bisogno, T., De Petrocellis, L., 1998. Endocannabinoids:
967 Endogenous cannabinoid receptor ligands with neuromodulatory action. *Trends*
968 *Neurosci.* 21, 521-528. [https://doi.org/10.1016/S0166-2236\(98\)01283-1](https://doi.org/10.1016/S0166-2236(98)01283-1)

969 Dincheva, I., Drysdale, A.T., Hartley, C.A., Johnson, D.C., Jing, D., King, E.C., Ra, S.,
970 Gray, J.M., Yang, R., DeGruccio, A.M., Huang, C., Cravatt, B.F., Glatt, C.E., Hill,
971 M.N., Casey, B.J., Lee, F.S., 2015. FAAH genetic variation enhances fronto-
972 amygdala function in mouse and human. *Nat. Commun.* 6.
973 <https://doi.org/10.1038/ncomms7395>

974 Dinh, T.P., Carpenter, D., Leslie, F.M., Freund, T.F., Katona, I., Sensi, S.L., Kathuria,
975 S., Piomelli, D., 2002. Brain monoglyceride lipase participating in
976 endocannabinoid inactivation. *Proc. Natl. Acad. Sci. U. S. A.* 99, 10819–10824.
977 <https://doi.org/10.1073/pnas.152334899>

- 978 Dinu, A.R., Rogobete, A.F., Bratu, T., Popovici, S.E., Bedreag, O.H., Papurica, M.,
979 Bratu, L.M., Sandesc, D., 2020. Cannabis Sativa Revisited—Crosstalk between
980 microRNA Expression, Inflammation, Oxidative Stress, and Endocannabinoid
981 Response System in Critically Ill Patients with Sepsis. *Cells* 9, 307.
982 <https://doi.org/10.3390/cells9020307>
- 983 Donvito, G., Nass, S.R., Wilkerson, J.L., Curry, Z.A., Schurman, L.D., Kinsey, S.G.,
984 Lichtman, A.H., 2018. The Endogenous Cannabinoid System: A Budding Source
985 of Targets for Treating Inflammatory and Neuropathic Pain.
986 *Neuropsychopharmacology* 43, 52–79. <https://doi.org/10.1038/npp.2017.204>
- 987 Du, H., Chen, X., Zhang, J., Chen, C., 2011. Inhibition of COX-2 expression by
988 endocannabinoid 2-arachidonoylglycerol is mediated via PPAR-g. *Br. J.*
989 *Pharmacol.* 163, 1533. <https://doi.org/10.1111/bph.2011.163.issue-7>
- 990 Ehrhart, J., Obregon, D., Mori, T., Hou, H., Sun, N., Bai, Y., Klein, T., Fernandez, F.,
991 Tan, J., Douglas Shytle, R., 2005. Stimulation of cannabinoid receptor 2 (CB2)
992 suppresses microglial activation. 58, 118-129. [https://doi.org/10.1186/1742-2094-](https://doi.org/10.1186/1742-2094-2-29)
993 [2-29](https://doi.org/10.1186/1742-2094-2-29)
- 994 Erickson, E.K., Grantham, E.K., Warden, A.S., Harris, R.A., 2019. Neuroimmune
995 signaling in alcohol use disorder. *Pharmacol. Biochem. Behav.* 177, 34–60.
996 <https://doi.org/10.1016/j.pbb.2018.12.007>
- 997 Fakhfouri, G., Ahmadiani, A., Rahimian, R., Grolla, A.A., Moradi, F., Haeri, A., 2012.
998 WIN55212-2 attenuates amyloid-beta-induced neuroinflammation in rats through
999 activation of cannabinoid receptors and PPAR-g pathway. *Neuropharmacology* 63,
1000 653–666. <https://doi.org/10.1016/j.neuropharm.2012.05.013>
- 1001 Fernandez-Lizarbe, S., Pascual, M., Guerri, C., 2009. Critical role of TLR4 response in
1002 the activation of microglia induced by ethanol. *J. Immunol.* 183, 4733–44.
1003 <https://doi.org/10.4049/jimmunol.0803590>
- 1004 Ferrier, L., Bérard, F., Debrauwer, L., Chabo, C., Langella, P., Buéno, L., Fioramonti,
1005 J., 2006. Impairment of the intestinal barrier by ethanol involves enteric microflora
1006 and mast cell activation in rodents. *Am. J. Pathol.* 168, 1148–1154.
1007 <https://doi.org/10.2353/ajpath.2006.050617>

- 1008 Franco, R., Fernández-Suárez, D., 2015. Alternatively activated microglia and
1009 macrophages in the central nervous system. *Prog. Neurobiol.* 131, 65-86.
1010 <https://doi.org/10.1016/j.pneurobio.2015.05.003>
- 1011 Freund, A.F., Katona, A., Piomelli, D., 2003. Role of Endogenous Cannabinoids in
1012 Synaptic Signaling. 83,1017-1066. <https://doi.org/10.1152/physrev.00004.2003.->
1013 Research
- 1014 Frontera, J.L., Gonzalez Pini, V.M., Messori, F.L., Brusco, A., 2018. Exposure to
1015 cannabinoid agonist WIN 55,212-2 during early adolescence increases alcohol
1016 preference and anxiety in CD1 mice. *Neuropharmacology* 137, 268–274.
1017 <https://doi.org/10.1016/j.neuropharm.2018.05.018>
- 1018 Fumagalli, M., Lombardi, M., Gressens, P., Verderio, C., 2018. How to reprogram
1019 microglia toward beneficial functions. 66, 2531-2549. *Glia*.
1020 <https://doi.org/10.1002/glia.23484>
- 1021 Galán-Ganga, M., del Río, R., Jiménez-Moreno, N., Díaz-Guerra, M., Lastres-Becker,
1022 I., 2020. Cannabinoid CB2 Receptor Modulation by the Transcription Factor NRF2
1023 is Specific in Microglial Cells. *Cell. Mol. Neurobiol.* 40, 167–177.
1024 <https://doi.org/10.1007/s10571-019-00719-y>
- 1025 Gao, B., Seki, E., Brenner, D.A., Friedman, S., Cohen, J.I., Nagy, L., Szabo, G.,
1026 Zakhari, S., 2011. Innate immunity in alcoholic liver disease. *Am. J. Physiol. -*
1027 *Gastrointest. Liver Physiol.* 300, 516-525.
1028 <https://doi.org/10.1152/ajpgi.00537.2010>
- 1029 Gilhus, N.E., Deuschl, G., 2019. Neuroinflammation — a common thread in
1030 neurological disorders. *Nat. Rev. Neurol.* <https://doi.org/10.1038/s41582-019->
1031 0227-8
- 1032 González-Reimers, E., Fernández-Rodríguez, C.M., Santolaria-Fernández, F., Vega-
1033 Prieto, M.J. de la, Martín-González, C., Gómez-Rodríguez, M.Á., Alemán-Valls,
1034 M.R., Rodríguez-Gaspar, M., 2011. Interleukin-15 and Other Myokines in Chronic
1035 Alcoholics. *Alcohol Alcohol.* 46, 529–533. <https://doi.org/10.1093/alcalc/agr064>
- 1036 Grabner, G.F., Zimmermann, R., Schicho, R., Taschler, U., 2017. Monoglyceride lipase

- 1037 as a drug target: At the crossroads of arachidonic acid metabolism and
1038 endocannabinoid signaling. *Pharmacol. Ther.* 175, 35–46.
1039 <https://doi.org/10.1016/j.pharmthera.2017.02.033>
- 1040 Greenhalgh, A.D., David, S., Bennett, F.C., 2020. Immune cell regulation of glia during
1041 CNS injury and disease. *Nat. Rev. Neurosci.* 21, 139-152.
1042 <https://doi.org/10.1038/s41583-020-0263-9>
- 1043 Guerri, C., Pascual, M., 2019. Impact of neuroimmune activation induced by alcohol or
1044 drug abuse on adolescent brain development. *Int. J. Dev. Neurosci.* 77, 89–98.
1045 <https://doi.org/10.1016/j.ijdevneu.2018.11.006>
- 1046 Guo, Y., Hong, W., Wang, X., Zhang, P., Körner, H., Tu, J., Wei, W., 2019.
1047 MicroRNAs in microglia: How do MicroRNAs affect activation, inflammation,
1048 polarization of microglia and mediate the interaction between microglia and
1049 glioma? *Front. Mol. Neurosci.* <https://doi.org/10.3389/fnmol.2019.00125>
- 1050 Hamelink, C., Hampson, A., Wink, D.A., Eiden, L.E., Eskay, R.L., 2005. Comparison
1051 of Cannabidiol, Antioxidants, and Diuretics in Reversing Binge Ethanol-Induced
1052 Neurotoxicity. *J. pharmacol. Exp. Ther.* 314, 780–788.
1053 <https://doi.org/10.1124/jpet.105.085779>
- 1054 Hansen, H.S., Rosenkilde, M.M., Holst, J.J., Schwartz, T.W., 2012. GPR119 as a fat
1055 sensor. *Trends Pharmacol. Sci.* 33, 374-381.
1056 <https://doi.org/10.1016/j.tips.2012.03.014>
- 1057 He, J., Crews, F.T., 2008. Increased MCP-1 and microglia in various regions of the
1058 human alcoholic brain. *Exp. Neurol.* 210, 349–358.
1059 <https://doi.org/10.1016/j.expneurol.2007.11.017>
- 1060 Heberlein, A., Käser, M., Lichtinghagen, R., Rhein, M., Lenz, B., Kornhuber, J., Bleich,
1061 S., Hillemecher, T., 2014. TNF- α and IL-6 serum levels: Neurobiological markers
1062 of alcohol consumption in alcohol-dependent patients? *Alcohol* 48, 671–676.
1063 <https://doi.org/10.1016/j.alcohol.2014.08.003>
- 1064 Huang, S.C.C., Everts, B., Ivanova, Y., O’Sullivan, D., Nascimento, M., Smith, A.M.,
1065 Beatty, W., Love-Gregory, L., Lam, W.Y., O’Neill, C.M., Yan, C., Du, H.,

- 1066 Abumrad, N.A., Urban, J.F., Artyomov, M.N., Pearce, E.L., Pearce, E.J., 2014.
1067 Cell-intrinsic lysosomal lipolysis is essential for alternative activation of
1068 macrophages. *Nat. Immunol.* 15, 846–855. <https://doi.org/10.1038/ni.2956>
- 1069 Ignacio, C., Hicks, S.D., Burke, P., Lewis, L., Szombathyne-Meszaros, Z., Middleton,
1070 F.A., 2015. Alterations in serum microRNA in humans with alcohol use disorders
1071 impact cell proliferation and cell death pathways and predict structural and
1072 functional changes in brain. *BMC Neurosci.* 16, 55.
1073 <https://doi.org/10.1186/s12868-015-0195-x>
- 1074 Irving, A.J., McDonald, N.A., Harkany, T., 2008. CB1 cannabinoid receptors:
1075 Molecular biology, second messenger coupling and polarized trafficking in
1076 neurons. *Cannabinoids and the Brain.* Springer US. 59–73.
1077 https://doi.org/10.1007/978-0-387-74349-3_5
- 1078 Jeon, Y.J., Yang, K.H., Pulaski, J.T., Kaminski, N.E., 1996. Attenuation of inducible
1079 nitric oxide synthase gene expression by Δ^9 - tetrahydrocannabinol is mediated
1080 through the inhibition of nuclear factor- κ B/Rel activation. *Mol. Pharmacol.* 50,
1081 334–341.
- 1082 Johns, D.G., Behm, D.J., Walker, D.J., Ao, Z., Shapland, E.M., Daniels, D.A., Riddick,
1083 M., Dowell, S., Staton, P.C., Green, P., Shabon, U., Bao, W., Aiyar, N., Yue, T.-L.,
1084 Brown, A.J., Douglas, S.A., 2007. The novel endocannabinoid receptor GPR55 is
1085 activated by atypical cannabinoids but does not mediate their vasodilator effects.
1086 *Br. J. Pharmacol.* 152, 825–831. <https://doi.org/10.1038/sj.bjp.0707419>
- 1087 Juknat, A., Gao, F., Coppola, G., Vogel, Z., Kozela, E., 2019. miRNA expression
1088 profiles and molecular networks in resting and LPS-activated BV-2 microglia—
1089 Effect of cannabinoids. *PLoS One* 14, e0212039.
1090 <https://doi.org/10.1371/journal.pone.0212039>
- 1091 Kane, C.J.M., Phelan, K.D., Douglas, J.C., Wagoner, G., Johnson, J.W., Xu, J., Phelan,
1092 P.S., Drew, P.D., 2014. Effects of Ethanol on Immune Response in the Brain:
1093 Region-Specific Changes in Adolescent Versus Adult Mice. *Alcohol. Clin. Exp.*
1094 *Res.* 38, 384–391. <https://doi.org/10.1111/acer.12244>
- 1095 Kaplan, B.L.F., 2013. The role of CB1 in immune modulation by cannabinoids.

- 1096 Pharmacol. Ther. 137, 365-374. <https://doi.org/10.1016/j.pharmthera.2012.12.004>
- 1097 Kaplan, J.S., Stella, N., Catterall, W.A., Westenbroek, R.E., 2017. Cannabidiol
1098 attenuates seizures and social deficits in a mouse model of Dravet syndrome. Proc.
1099 Natl. Acad. Sci. 114, 11229–11234. <https://doi.org/10.1073/pnas.1711351114>
- 1100 Kapoor, M., Wang, J.C., Farris, S.P., Liu, Y., McClintick, J., Gupta, I., Meyers, J.L.,
1101 Bertelsen, S., Chao, M., Nurnberger, J., Tischfield, J., Harari, O., Zeran, L.,
1102 Hesselbrock, V., Bauer, L., Raj, T., Porjesz, B., Agrawal, A., Foroud, T.,
1103 Edenberg, H.J., Mayfield, R.D., Goate, A., 2019. Analysis of whole genome-
1104 transcriptomic organization in brain to identify genes associated with alcoholism.
1105 Transl. Psychiatry 9, 1–11. <https://doi.org/10.1038/s41398-019-0384-y>
- 1106 Karoly, H.C., Bidwell, L.C., Mueller, R.L., Hutchison, K.E., 2018. Investigating the
1107 Relationships Between Alcohol Consumption, Cannabis Use, and Circulating
1108 Cytokines: A Preliminary Analysis. Alcohol. Clin. Exp. Res. 42, 531–539.
1109 <https://doi.org/10.1111/acer.13592>
- 1110 Katona, I., ta Sperlá gh, B., Sík, A., Kä falvi, A., Sylvester Vizi, E., Mackie, K., Freund,
1111 T.F., 1999. Presynaptically Located CB1 Cannabinoid Receptors Regulate GABA
1112 Release from Axon Terminals of Specific Hippocampal Interneurons. J.
1113 Neurosci.19, 4544-4558. <https://doi.org/10.1523/JNEUROSCI.19-11-04544.1999>
- 1114 Katona, I., Urbán, G.M., Wallace, M., Ledent, C., Jung, K.M., Piomelli, D., Mackie, K.,
1115 Freund, T.F., 2006. Molecular composition of the endocannabinoid system at
1116 glutamatergic synapses. J. Neurosci. 26, 5628–5637.
1117 <https://doi.org/10.1523/JNEUROSCI.0309-06.2006>
- 1118 Kelly, R., Joers, V., Tansey, M.G., McKernan, D.P., Dowd, E., 2020. Microglial
1119 Phenotypes and Their Relationship to the Cannabinoid System: Therapeutic
1120 Implications for Parkinson’s Disease. Molecules 25, 453.
1121 <https://doi.org/10.3390/molecules25030453>
- 1122 Kong, W.-L., Peng, Y.-Y., Peng, B.-W., 2017. Modulation of neuroinflammation: Role
1123 and therapeutic potential of TRPV1 in the neuro-immune axis. Brain. Behav.
1124 Immun. 64, 354-366. <https://doi.org/10.1016/j.bbi.2017.03.007>

- 1125 Kozela, E., Pietr, M., Juknat, A., Rimmerman, N., Levy, R., Vogel, Z., 2010.
1126 Cannabinoids Δ^9 -Tetrahydrocannabinol and Cannabidiol Differentially Inhibit
1127 the Lipopolysaccharide-activated NF- κ B and Interferon- β /STAT Proinflammatory
1128 Pathways in BV-2 Microglial Cells. *J. Biol. Chem.* 285, 1616–1626.
1129 <https://doi.org/10.1074/jbc.M109.069294>
- 1130 Kyzar, E.J., Floreani, C., Teppen, T.L., Pandey, S.C., 2016. Adolescent Alcohol
1131 Exposure: Burden of Epigenetic Reprogramming, Synaptic Remodeling, and Adult
1132 Psychopathology. *Front. Neurosci.* 10. <https://doi.org/10.3389/fnins.2016.00222>
- 1133 Lamont, M.G., McCallum, P., Head, N., Blundell, J., Weber, J.T., 2020. Binge drinking
1134 in male adolescent rats and its relationship to persistent behavioral impairments
1135 and elevated proinflammatory/proapoptotic proteins in the cerebellum.
1136 *Psychopharmacology (Berl)*. 23, 1305-1315. [https://doi.org/10.1007/s00213-020-](https://doi.org/10.1007/s00213-020-05458-3)
1137 [05458-3](https://doi.org/10.1007/s00213-020-05458-3)
- 1138 Lauckner, J.E., Jensen, J.B., Chen, H.Y., Lu, H.C., Hille, B., Mackie, K., 2008. GPR55
1139 is a cannabinoid receptor that increases intracellular calcium and inhibits M
1140 current. *Proc. Natl. Acad. Sci. U. S. A.* 105, 2699–2704.
1141 <https://doi.org/10.1073/pnas.0711278105>
- 1142 Leclercq, S., Cani, P.D., Neyrinck, A.M., Stärkel, P., Jamar, F., Mikolajczak, M.,
1143 Delzenne, N.M., De Timary, P., 2012. Role of intestinal permeability and
1144 inflammation in the biological and behavioral control of alcohol-dependent
1145 subjects. *Brain. Behav. Immun.* 26, 911–918.
1146 <https://doi.org/10.1016/j.bbi.2012.04.001>
- 1147 Leclercq, S., De Saeger, C., Delzenne, N., de Timary, P., Stärkel, P., 2014. Role of
1148 inflammatory pathways, blood mononuclear cells, and gut-derived bacterial
1149 products in alcohol dependence. *Biol. Psychiatry* 76, 725–33.
1150 <https://doi.org/10.1016/j.biopsych.2014.02.003>
- 1151 Lewohl, J.M., Wang, L., Miles, M.F., Zhang, L., Dodd, P.R., Harris, R.A., 2000. Gene
1152 expression in human alcoholism: microarray analysis of frontal cortex. *Alcohol.*
1153 *Clin. Exp. Res.* 24, 1873–82.
- 1154 Li, L., Wu, Y., Wang, Y., Wu, J., Song, L., Xian, W., Yuan, S., Pei, L., Shang, Y.,

- 1155 2014. Resolvin D1 promotes the interleukin-4-induced alternative activation in
1156 BV-2 microglial cells. *J. Neuroinflammation* 11, 72. [https://doi.org/10.1186/1742-](https://doi.org/10.1186/1742-2094-11-72)
1157 [2094-11-72](https://doi.org/10.1186/1742-2094-11-72)
- 1158 Liao, N.P.D., Laktyushin, A., Lucet, I.S., Murphy, J.M., Yao, S., Whitlock, E.,
1159 Callaghan, K., Nicola, N.A., Kershaw, N.J., Babon, J.J., 2018. The molecular basis
1160 of JAK/STAT inhibition by SOCS1. *Nat. Commun.* 9, 1–14.
1161 <https://doi.org/10.1038/s41467-018-04013-1>
- 1162 Lippai, D., Bala, S., Catalano, D., Kodys, K., Szabo, G., 2014. Micro-RNA-155
1163 Deficiency Prevents Alcohol-Induced Serum Endotoxin Increase and Small Bowel
1164 Inflammation in Mice. *Alcohol. Clin. Exp. Res.* 38, 2217–2224.
1165 <https://doi.org/10.1111/acer.12483>
- 1166 Lippai, D., Bala, S., Csak, T., Kurt-Jones, E.A., Szabo, G., 2013. Chronic Alcohol-
1167 Induced microRNA-155 Contributes to Neuroinflammation in a TLR4-Dependent
1168 Manner in Mice. *PLoS One* 8, 1–10. <https://doi.org/10.1371/journal.pone.0070945>
- 1169 Liput, D., Pauly, J., Stinchcomb, A., Nixon, K., 2017. Binge Alcohol Exposure
1170 Transiently Changes the Endocannabinoid System: A Potential Target to Prevent
1171 Alcohol-Induced Neurodegeneration. *Brain Sci.* 7, 158.
1172 <https://doi.org/10.3390/brainsci7120158>
- 1173 Liput, D.J., Hammell, D.C., Stinchcomb, A.L., Nixon, K., 2013. Transdermal Delivery
1174 of Cannabidiol Attenuates Binge Alcohol-Induced Neurodegeneration in a Rodent
1175 Model of an Alcohol Use Disorder. *Pharmacol Biochem Behav* 111, 120–127.
1176 <https://doi.org/10.1016/j.pbb.2013.08.013>
- 1177 Little, J., Villanueva, E., Klegeris, A., 2011. Therapeutic Potential of Cannabinoids in
1178 the Treatment of Neuroinflammation Associated with Parkinsons Disease. *Mini-*
1179 *Reviews Med. Chem.* 11, 582–590. <https://doi.org/10.2174/138955711795906905>
- 1180 Liu, B., Song, S., Jones, P.M., Persaud, S.J., 2015. GPR55: From orphan to metabolic
1181 regulator? *Pharmacol. Ther.* 145, 35-42
1182 <https://doi.org/10.1016/j.pharmthera.2014.06.007>
- 1183 Long, J.Z., Li, W., Booker, L., Burston, J.J., Kinsey, S.G., Schlosburg, J.E., Pavón, F.J.,

- 1184 Serrano, A.M., Selley, D.E., Parsons, L.H., Lichtman, A.H., Cravatt, B.F., Chem,
1185 N., Author, B., 2009. Selective blockade of 2-arachidonoylglycerol hydrolysis
1186 produces cannabinoid behavioral effects. *Nat Chem Biol* 5, 37–44.
1187 <https://doi.org/10.1038/nchembio.129>
- 1188 Lou, Z.-Y., Yu, W.-B., Chen, J., Li, L., Jiang, L.-S., Xiao, B.-G., Liu, Z.-G., 2016.
1189 Neuroprotective Effect is Driven Through the Upregulation of CB1 Receptor in
1190 Experimental Autoimmune Encephalomyelitis. *J. Mol. Neurosci.* 58, 193–200.
1191 <https://doi.org/10.1007/s12031-015-0656-9>
- 1192 Lowin, T., Straub, R.H., 2015. Cannabinoid-based drugs targeting CB1 and TRPV1, the
1193 sympathetic nervous system, and arthritis. *Arthritis Res. Ther.*
1194 <https://doi.org/10.1186/s13075-015-0743-x>
- 1195 Lu, H.C., MacKie, K., 2016. An introduction to the endogenous cannabinoid system.
1196 *Biol. Psychiatry* 79, 516–525. <https://doi.org/10.1016/j.biopsych.2015.07.028>
- 1197 Mandal, C., Halder, D., Jung, K.H., Chai, Y.G., 2018. Maternal alcohol consumption
1198 and altered miRNAs in the developing fetus: Context and future perspectives. *J.*
1199 *Appl. Toxicol.* 38, 100–107. <https://doi.org/10.1002/jat.3504>
- 1200 Maroof, N., Pardon, M.C., Kendall, D.A., 2013. Endocannabinoid signalling in
1201 Alzheimer’s disease. *Biochemical Society Transactions*. Portland Press. 1583–
1202 1587. <https://doi.org/10.1042/BST20130140>
- 1203 Maroso, M., Balosso, S., Ravizza, T., Liu, J., Bianchi, M.E., Vezzani, A., 2011.
1204 Interleukin-1 type 1 receptor/Toll-like receptor signalling in epilepsy: The
1205 importance of IL-1beta and high-mobility group box 1. *Journal of Internal*
1206 *Medicine.* 319–326. <https://doi.org/10.1111/j.1365-2796.2011.02431.x>
- 1207 Marrone, M.C., Morabito, A., Giustizieri, M., Chiurchiù, V., Leuti, A., Mattioli, M.,
1208 Marinelli, Sara, Riganti, L., Lombardi, M., Murana, E., Totaro, A., Piomelli, D.,
1209 Ragozzino, D., Oddi, S., Maccarrone, M., Verderio, C., Marinelli, Silvia, 2017.
1210 TRPV1 channels are critical brain inflammation detectors and neuropathic pain
1211 biomarkers in mice. *Nat. Commun.* 8. <https://doi.org/10.1038/ncomms15292>
- 1212 Marsicano, G., 2003. CB1 Cannabinoid Receptors and On-Demand Defense Against

- 1213 Excitotoxicity. *Science* 302, 84–88. <https://doi.org/10.1126/science.1088208>
- 1214 Matsuda, L.A., Lolait, S.J., Brownstein, M.J., Young, A.C., Bonner, T.I., 1990.
1215 Structure of a cannabinoid receptor and functional expression of the cloned cDNA.
1216 *Nature* 346, 561–564. <https://doi.org/10.1038/346561a0>
- 1217 McClain, J.A., Morris, S.A., Deeny, M.A., Marshall, S.A., Hayes, D.M., Kiser, Z.M.,
1218 Nixon, K., 2011. Adolescent binge alcohol exposure induces long-lasting partial
1219 activation of microglia. *Brain. Behav. Immun.* 25, S120–S128.
1220 <https://doi.org/10.1016/j.bbi.2011.01.006>
- 1221 McHugh, D., 2012. GPR18 in microglia: Implications for the CNS and
1222 endocannabinoid system signalling. *Br. J. Pharmacol.* 167, 1575-1582.
1223 <https://doi.org/10.1111/j.1476-5381.2012.02019.x>
- 1224 Mecha, M., Carrillo-Salinas, F.J., Feliú, A., Mestre, L., Guaza, C., 2016. Microglia
1225 activation states and cannabinoid system: Therapeutic implications. *Pharmacol.*
1226 *Ther.* 166, 40–55. <https://doi.org/10.1016/j.pharmthera.2016.06.011>
- 1227 Melbourne, J.K., Thompson, K.R., Peng, H., Nixon, K., 2019. Its complicated: The
1228 relationship between alcohol and microglia in the search for novel
1229 pharmacotherapeutic targets for alcohol use disorders. *Prog. Mol. Biol. Transl. Sci.*
1230 167, 179–221. <https://doi.org/10.1016/BS.PMBTS.2019.06.011>
- 1231 Moreno, S., Farioli-Vecchioli, S., Ceru`b, A.M.P., Ceru`b, C., 2004.
1232 Immunolocalization of peroxisome proliferator-activated receptors and retinoid X
1233 receptors in the adult rat CNS. *Neuroscience* 123, 131–145.
1234 <https://doi.org/10.1016/j.neuroscience.2003.08.064>
- 1235 Muller, C., Morales, P., Reggio, P.H., 2018. Cannabinoid Ligands Targeting TRP
1236 Channels. *Front. Mol. Neurosci.* 11, 487.
1237 <https://doi.org/10.3389/fnmol.2018.00487>
- 1238 Munro, S., Thomas, K.L., Abu-Shaar, M., 1993. Molecular characterization of a
1239 peripheral receptor for cannabinoids. *Nature* 365, 61–65.
1240 <https://doi.org/10.1038/365061a0>

- 1241 Murray, P.J., 2007. The JAK-STAT Signaling Pathway: Input and Output Integration. *J.*
1242 *Immunol.* 178, 2623–2629. <https://doi.org/10.4049/jimmunol.178.5.2623>
- 1243 Murray, P.J., 2006. STAT3-mediated anti-inflammatory signalling. *Biochem. Soc.*
1244 *Trans.* 34, 1028–1031. <https://doi.org/10.1042/BST0341028>
- 1245 Nabavi, S.M., Ahmed, T., Nawaz, M., Devi, K.P., Balan, D.J., Pittalà, V., Argüelles-
1246 Castilla, S., Testai, L., Khan, H., Sureda, A., de Oliveira, M.R., Vacca, R.A., Xu,
1247 S., Yousefi, B., Curti, V., Daglia, M., Sobarzo-Sánchez, E., Filosa, R., Nabavi,
1248 S.F., Majidinia, M., Dehpour, A.R., Shirooie, S., 2019. Targeting STATs in
1249 neuroinflammation: The road less traveled! *Pharmacol. Res.* 141,73-84.
1250 <https://doi.org/10.1016/j.phrs.2018.12.004>
- 1251 Nair, M.P., Figueroa, G., Casteleiro, G., Muñoz, K., Agudelo, M., 2015. Alcohol Versus
1252 Cannabinoids: A Review of Their Opposite Neuro-Immunomodulatory Effects and
1253 Future Therapeutic Potentials. *J. Alcohol. drug Depend.* 3.
1254 <https://doi.org/10.4172/2329-6488.1000184>
- 1255 Nomura, D.K., Morrison, B.E., Blankman, J.L., Long, J.Z., Kinsey, S.G., Marcondes,
1256 M.C.G., Ward, A.M., Hahn, Y.K., Lichtman, A.H., Conti, B., Cravatt, B.F., 2011.
1257 Endocannabinoid hydrolysis generates brain prostaglandins that promote
1258 neuroinflammation. *Science.* 334, 809–813.
1259 <https://doi.org/10.1126/science.1209200>
- 1260 O’Sullivan, S.E., 2016. An update on PPAR activation by cannabinoids. *Br. J.*
1261 *Pharmacol.* 173, 1899-1910. <https://doi.org/10.1111/bph.13497>
- 1262 Okamoto, Y., Morishita, J., Tsuboi, K., Tonai, T., Ueda, N., 2004. Molecular
1263 characterization of a phospholipase D generating anandamide and its congeners. *J.*
1264 *Biol. Chem.* 279, 5298–305. <https://doi.org/10.1074/jbc.M306642200>
- 1265 Orio, L., Alen, F., Pavón, F.J., Serrano, A., García-Bueno, B., 2019.
1266 Oleoylethanolamide, neuroinflammation, and alcohol abuse. *Front. Mol. Neurosci.*
1267 <https://doi.org/10.3389/fnmol.2018.00490>
- 1268 Park, J.S., Svetkauskaite, D., He, Q., Kim, J.Y., Strassheim, D., Ishizaka, A., Abraham,
1269 E., 2004. Involvement of Toll-like Receptors 2 and 4 in Cellular Activation by

- 1270 High Mobility Group Box 1 Protein. *J. Biol. Chem.* 279, 7370–7377.
1271 <https://doi.org/10.1074/jbc.M306793200>
- 1272 Pascual, M., Blanco, A.M., Cauli, O., Miñarro, J., Guerri, C., 2007. Intermittent ethanol
1273 exposure induces inflammatory brain damage and causes long-term behavioural
1274 alterations in adolescent rats. *Eur. J. Neurosci.* 25, 541–550.
1275 <https://doi.org/10.1111/j.1460-9568.2006.05298.x>
- 1276 Pascual, M., Montesinos, J., Guerri, C., 2018. Role of the innate immune system in the
1277 neuropathological consequences induced by adolescent binge drinking. *J.*
1278 *Neurosci. Res.* 96, 765–780. <https://doi.org/10.1002/jnr.24203>
- 1279 Pelicão, R., Santos, M.C., Freitas-Lima, L.C., Meyrelles, S.S., Vasquez, E.C.,
1280 Nakamura-Palacios, E.M., Rodrigues, L.C.M., 2016. URB597 inhibits oxidative
1281 stress induced by alcohol binge in the prefrontal cortex of adolescent rats.
1282 *Neurosci. Lett.* 624, 17–22. <https://doi.org/10.1016/j.neulet.2016.04.068>
- 1283 Peñasco, S., Rico-Barrio, I., Puente, N., Fontaine, C.J., Ramos, A., Reguero, L.,
1284 Gerrikagoitia, I., de Fonseca, F.R., Suarez, J., Barrondo, S., Aretxabala, X., García
1285 del Caño, G., Sallés, J., Elezgarai, I., Nahirney, P.C., Christie, B.R., Grandes, P.,
1286 2020. Intermittent ethanol exposure during adolescence impairs cannabinoid type 1
1287 receptor-dependent long-term depression and recognition memory in adult mice.
1288 *Neuropsychopharmacology* 45, 309–318. [https://doi.org/10.1038/s41386-019-](https://doi.org/10.1038/s41386-019-0530-5)
1289 [0530-5](https://doi.org/10.1038/s41386-019-0530-5)
- 1290 Perkins, A.E., Varlinskaya, E.I., Deak, T., 2019. From adolescence to late aging: A
1291 comprehensive review of social behavior, alcohol, and neuroinflammation across
1292 the lifespan. *International Review of Neurobiology.* Academic Press Inc. 148, 231–
1293 303. <https://doi.org/10.1016/bs.irn.2019.08.001>
- 1294 Petrosino, S., Schiano Moriello, A., Cerrato, S., Fusco, M., Puigdemont, A., De
1295 Petrocellis, L., Di Marzo, V., 2016. The anti-inflammatory mediator
1296 palmitoylethanolamide enhances the levels of 2-arachidonoyl-glycerol and
1297 potentiates its actions at TRPV1 cation channels. *Br. J. Pharmacol.* 173, 1154–
1298 1162. <https://doi.org/10.1111/bph.13084>
- 1299 Pietr, M., Kozela, E., Levy, R., Rimmerman, N., Lin, Y.H., Stella, N., Vogel, Z., Juknat,

- 1300 A., 2009. Differential changes in GPR55 during microglial cell activation. *FEBS*
1301 *Lett.* 583, 2071–2076. <https://doi.org/10.1016/j.febslet.2009.05.028>
- 1302 Qin, L., Crews, F.T., 2012. NADPH oxidase and reactive oxygen species contribute to
1303 alcohol-induced microglial activation and neurodegeneration. *J.*
1304 *Neuroinflammation* 9, 5. <https://doi.org/10.1186/1742-2094-9-5>
- 1305 Qin, L., He, J., Hanes, R.N., Pluzarev, O., Hong, J.S., Crews, F.T., 2008. Increased
1306 systemic and brain cytokine production and neuroinflammation by endotoxin
1307 following ethanol treatment. *J. Neuroinflammation* 5. [https://doi.org/10.1186/1742-](https://doi.org/10.1186/1742-2094-5-10)
1308 [2094-5-10](https://doi.org/10.1186/1742-2094-5-10)
- 1309 Rehm, J., Shield, K.D., 2019. Global Burden of Alcohol Use Disorders and Alcohol
1310 Liver Disease. *Biomedicines* 7, 99. <https://doi.org/10.3390/biomedicines7040099>
- 1311 Ribeiro, R., Wen, J., Li, S., Zhang, Y., 2013. Involvement of ERK1/2, cPLA 2 and NF-
1312 kB in microglia suppression by cannabinoid receptor agonists and antagonists.
1313 *Prostaglandins Other Lipid Mediat.* 100, 1–14.
1314 <https://doi.org/10.1016/j.prostaglandins.2012.11.003>
- 1315 Rivera, P., Fernández-Arjona, M. del M., Silva-Peña, D., Blanco, E., Vargas, A., López-
1316 Ávalos, M.D., Grondona, J.M., Serrano, A., Pavón, F.J., Rodríguez de Fonseca, F.,
1317 Suárez, J., 2018. Pharmacological blockade of fatty acid amide hydrolase (FAAH)
1318 by URB597 improves memory and changes the phenotype of hippocampal
1319 microglia despite ethanol exposure. *Biochem. Pharmacol.* 157, 244–257.
1320 <https://doi.org/10.1016/j.bcp.2018.08.005>
- 1321 Rivera, P., Silva-Peña, D., Blanco, E., Vargas, A., Arrabal, S., Serrano, A., Pavón, F.J.,
1322 Bindila, L., Lutz, B., Rodríguez de Fonseca, F., Suárez, J., 2019.
1323 Oleoylethanolamide restores alcohol-induced inhibition of neuronal proliferation
1324 and microglial activity in striatum. *Neuropharmacology* 146, 184–197.
1325 <https://doi.org/10.1016/j.neuropharm.2018.11.037>
- 1326 Robinson, R.H., Meissler, J.J., Fan, X., Yu, D., Adler, M.W., Eisenstein, T.K., 2015. A
1327 CB2-Selective Cannabinoid Suppresses T-Cell Activities and Increases Tregs and
1328 IL-10. *J. Neuroimmune Pharmacol.* 10, 318–332. [https://doi.org/10.1007/s11481-](https://doi.org/10.1007/s11481-015-9611-3)
1329 [015-9611-3](https://doi.org/10.1007/s11481-015-9611-3)

- 1330 Rodrigues, L.C.M.M., Gobira, P.H., de Oliveira, A.C., Pelicao, R., Teixeira, A.L.,
1331 Moreira, F.A., Campos, A.C., Pelicão, R., Teixeira, A.L., Moreira, F.A., Campos,
1332 A.C., 2014. Neuroinflammation as a possible link between cannabinoids and
1333 addiction. *Acta Neuropsychiatr.* 26, 334–346. <https://doi.org/10.1017/neu.2014.24>
- 1334 Rojo, A.I., Pajares, M., García-Yagüe, A.J., Buendia, I., Van Leuven, F., Yamamoto,
1335 M., López, M.G., Cuadrado, A., 2018. Deficiency in the transcription factor NRF2
1336 worsens inflammatory parameters in a mouse model with combined tauopathy and
1337 amyloidopathy. *Redox Biol.* 18, 173–180.
1338 <https://doi.org/10.1016/j.redox.2018.07.006>
- 1339 Ryberg, E., Larsson, N., Sjögren, S., Hjorth, S., Hermansson, N.-O., Leonova, J.,
1340 Elebring, T., Nilsson, K., Drmota, T., Greasley, P.J., 2007. The orphan receptor
1341 GPR55 is a novel cannabinoid receptor. *Br. J. Pharmacol.* 152, 1092–1101.
1342 <https://doi.org/10.1038/sj.bjp.0707460>
- 1343 Saijo, K., Glass, C.K., 2011. Microglial cell origin and phenotypes in health and
1344 disease. *Nat. Rev. Immunol.* 11, 775–787. <https://doi.org/10.1038/nri3086>
- 1345 Sanchez-Alavez, M., Nguyen, W., Mori, S., Wills, D.N., Otero, D., Ehlers, C.L., Conti,
1346 B., 2019. Time course of microglia activation and brain and blood
1347 cytokine/chemokine levels following chronic ethanol exposure and protracted
1348 withdrawal in rats. *Alcohol* 76, 37–45.
1349 <https://doi.org/10.1016/j.alcohol.2018.07.005>
- 1350 Sanchez-Marin, L., Pavon, F.J., Decara, J., Suarez, J., Gavito, A., Castilla-Ortega, E.,
1351 Rodriguez de Fonseca, F., Serrano, A., 2017. Effects of Intermittent Alcohol
1352 Exposure on Emotion and Cognition: A Potential Role for the Endogenous
1353 Cannabinoid System and Neuroinflammation. *Front. Behav. Neurosci.* 11, 15.
1354 <https://doi.org/10.3389/fnbeh.2017.00015>
- 1355 Schlosburg, J.E., Blankman, J.L., Long, J.Z., Nomura, D.K., Pan, B., Kinsey, S.G.,
1356 Nguyen, P.T., Ramesh, D., Booker, L., Burston, J.J., Thomas, E.A., Selley, D.E.,
1357 Sim-Selley, L.J., Liu, Q., Lichtman, A.H., Cravatt, B.F., 2010. Chronic
1358 monoacylglycerol lipase blockade causes functional antagonism of the
1359 endocannabinoid system. *Nat Neurosci* 13, 1113–1119.

- 1360 <https://doi.org/10.1038/nm.2616>
- 1361 Sheng, W.S., Hu, S., Min, X., Cabral, G.A., Lokensgard, J.R., Peterson, P.K., 2005.
1362 Synthetic cannabinoid WIN55,212-2 inhibits generation of inflammatory
1363 mediators by IL-1B-stimulated human astrocytes. *Glia* 49, 211–219.
1364 <https://doi.org/10.1002/glia.20108>
- 1365 Silva-Peña, D., Rivera, P., Alén, F., Vargas, A., Rubio, L., García-Marchena, N., Pavón,
1366 F.J., Serrano, A., Rodríguez de Fonseca, F., Suárez, J., 2019. Oleoylethanolamide
1367 Modulates BDNF-ERK Signaling and Neurogenesis in the Hippocampi of Rats
1368 Exposed to Δ^9 -THC and Ethanol Binge Drinking During Adolescence. *Front. Mol.*
1369 *Neurosci.* 12, 96. <https://doi.org/10.3389/fnmol.2019.00096>
- 1370 Smith, S.R., Terminelli, C., Denhardt, G., 2000. Effects of cannabinoid receptor agonist
1371 and antagonist ligands on production of inflammatory cytokines and anti-
1372 inflammatory interleukin-10 in endotoxemic mice. *J. Pharmacol. Exp. Ther.* 293,
1373 136–150.
- 1374 Soethoudt, M., Grether, U., Fingerle, J., Grim, T.W., Fezza, F., De Petrocellis, L.,
1375 Ullmer, C., Rothenhäusler, B., Perret, C., Van Gils, N., Finlay, D., Macdonald, C.,
1376 Chicca, A., Gens, M.D., Stuart, J., De Vries, H., Mastrangelo, N., Xia, L.,
1377 Alachouzos, G., Baggelaar, M.P., Martella, A., Mock, E.D., Deng, H., Heitman,
1378 L.H., Connor, M., Marzo, V. Di, Gertsch, J., Lichtman, A.H., Maccarrone, M.,
1379 Glass, M., Van Der Stelt, M., 2017. Cannabinoid CB 2 receptor ligand profiling
1380 reveals biased signalling and off-target activity. *Nat. Commun.*
1381 <https://doi.org/10.1038/ncomms13958>
- 1382 Stella, N., 2010. Cannabinoid and cannabinoid-like receptors in microglia, astrocytes,
1383 and astrocytomas. *Glia* 58, 1017–1030. <https://doi.org/10.1002/glia.20983>
- 1384 Vallée, A., Lecarpentier, Y., Guillevin, R., Vallée, J.-N., 2017. Effects of cannabidiol
1385 interactions with Wnt/ β -catenin pathway and PPAR γ on oxidative stress and
1386 neuroinflammation in Alzheimer's disease. *Acta Biochim. Biophys. Sin.*
1387 (Shanghai). 49, 853–866. <https://doi.org/10.1093/abbs/gmx073>
- 1388 Vallés, S.L., Blanco, A.M., Pascual, M., Guerri, C., 2004. Chronic ethanol treatment
1389 enhances inflammatory mediators and cell death in the brain and in astrocytes.

- 1390 Brain Pathol. 14, 365–71.
- 1391 Varga, T., Czimmerer, Z., Nagy, L., 2011a. PPARs are a unique set of fatty acid
1392 regulated transcription factors controlling both lipid metabolism and inflammation.
1393 Biochim. Biophys. Acta - Mol. Basis Dis. 1812, 1007–1022.
1394 <https://doi.org/10.1016/J.BBADIS.2011.02.014>
- 1395 Varga, T., Czimmerer, Z., Nagy, L., 2011b. PPARs are a unique set of fatty acid
1396 regulated transcription factors controlling both lipid metabolism and inflammation.
1397 Biochim. Biophys. Acta - Mol. Basis Dis. 1812, 1007–1022.
1398 <https://doi.org/10.1016/J.BBADIS.2011.02.014>
- 1399 Walter, L., Franklin, A., Witting, A., Wade, C., Xie, Y., Kunos, G., Mackie, K., Stella,
1400 N., 2003. Nonpsychotropic Cannabinoid Receptors Regulate Microglial Cell
1401 Migration.
- 1402 Wang, D., Shi, L., Xin, W., Xu, Jiancheng, Xu, Jing, Li, Q., Xu, Z., Wang, J., Wang,
1403 G., Yao, W., He, B., Yang, Y., Hu, M., 2017. Activation of PPAR γ inhibits pro-
1404 inflammatory cytokines production by upregulation of miR-124 in vitro and
1405 in vivo. Biochem. Biophys. Res. Commun. 486, 726–731.
1406 <https://doi.org/10.1016/j.bbrc.2017.03.106>
- 1407 Wang, Z., Yang, Y., Yang, H., Capó-Aponte, J.E., Tachado, S.D., Wolosin, J.M.,
1408 Reinach, P.S., 2011. NF- κ B feedback control of JNK1 activation modulates
1409 TRPV1-induced increases in IL-6 and IL-8 release by human corneal epithelial
1410 cells. Mol. Vis. 17, 3137–46.
- 1411 Wen, J., Ribeiro, R., Tanaka, M., Zhang, Y., 2015. Activation of CB2 receptor is
1412 required for the therapeutic effect of ABHD6 inhibition in experimental
1413 autoimmune encephalomyelitis. Neuropharmacology 99, 196–209.
1414 <https://doi.org/10.1016/j.neuropharm.2015.07.010>
- 1415 World Health Organization, W., 2018. Global status report on alcohol and health 2018.
1416 World Health Organization.
- 1417 Yamanaka, M., Ishikawa, T., Griep, A., Axt, D., Kummer, M.P., Heneka, M.T., 2012.
1418 PPAR γ /RXRA-induced and CD36-mediated microglial amyloid- β phagocytosis

- 1419 results in cognitive improvement in amyloid precursor protein/presenilin 1 mice. *J.*
1420 *Neurosci.* 32, 17321–17331. <https://doi.org/10.1523/JNEUROSCI.1569-12.2012>
- 1421 Yang, L., Guo, H., Li, Y., Meng, X., Yan, L., Dan Zhang, Wu, S., Zhou, H., Peng, L.,
1422 Xie, Q., Jin, X., 2016. Oleoylethanolamide exerts anti-inflammatory effects on
1423 LPS-induced THP-1 cells by enhancing PPAR α signaling and inhibiting the NF- κ B
1424 and ERK1/2/AP-1/STAT3 pathways. *Sci. Rep.* 6, 34611.
1425 <https://doi.org/10.1038/srep34611>
- 1426 Yang, Y., Yang, H., Wang, Z., Varadaraj, K., Kumari, S.S., Mergler, S., Okada, Y.,
1427 Saika, S., Kingsley, P.J., Marnett, L.J., Reinach, P.S., Edu, P., 2013. Cannabinoid
1428 receptor 1 suppresses transient receptor potential vanilloid 1-induced inflammatory
1429 responses to corneal injury. *Cell Signal* 25, 501–511.
1430 <https://doi.org/10.1016/j.cellsig.2012.10.015>
- 1431 Yuan, M., Kiertscher, S.M., Cheng, Q., Zoumalan, R., Tashkin, D.P., Roth, M.D., 2002.
1432 Δ 9-Tetrahydrocannabinol regulates Th1/Th2 cytokine balance in activated human
1433 T cells. *J. Neuroimmunol.* 133, 124–131. [https://doi.org/10.1016/S0165-](https://doi.org/10.1016/S0165-5728(02)00370-3)
1434 [5728\(02\)00370-3](https://doi.org/10.1016/S0165-5728(02)00370-3)
- 1435 Zarruk, J.G., Fernández-López, D., García-Yébenes, I., García-Gutiérrez, M.S.,
1436 Vivancos, J., Nombela, F., Torres, M., Burguete, M.C., Manzanares, J., Lizasoain,
1437 I., Moro, M.A., 2012. Cannabinoid type 2 receptor activation downregulates
1438 stroke-induced classic and alternative brain macrophage/microglial activation
1439 concomitant to neuroprotection. *Stroke* 43, 211–9.
1440 <https://doi.org/10.1161/STROKEAHA.111.631044>
- 1441 Zhang, M., An, C., Gao, Y., Leak, R.K., Chen, J., Zhang, F., 2013. Emerging roles of
1442 Nrf2 and phase II antioxidant enzymes in neuroprotection. *Prog. Neurobiol.* 100,
1443 30-47. <https://doi.org/10.1016/j.pneurobio.2012.09.003>
- 1444 Zhao, X., Sun, G., Ting, S.M., Song, S., Zhang, J., Edwards, N.J., Aronowski, J., 2015.
1445 Cleaning up after ICH: The role of Nrf2 in modulating microglia function and
1446 hematoma clearance. *J. Neurochem.* 133, 144–152.
1447 <https://doi.org/10.1111/jnc.12974>
- 1448 Zhou, Y., Huang, T., Lee, F., Kreek, M.J., 2016. Involvement of Endocannabinoids in

- 1449 Alcohol “Binge” Drinking: Studies of Mice with Human Fatty Acid Amide
1450 Hydrolase Genetic Variation and After CB1 Receptor Antagonists. *Alcohol. Clin.*
1451 *Exp. Res.* 40, 467–473. <https://doi.org/10.1111/acer.12989>
- 1452 Zhou, Y., Schwartz, B.I., Giza, J., Gross, S.S., Lee, F.S., Kreek, M.J., 2017. Blockade
1453 of alcohol escalation and “relapse” drinking by pharmacological FAAH inhibition
1454 in male and female C57BL/6J mice. *Psychopharmacology (Berl)*. 234, 2955–2970.
1455 <https://doi.org/10.1007/s00213-017-4691-9>
- 1456 Zou, J.Y., Crews, F.T., 2005. TNF α potentiates glutamate neurotoxicity by inhibiting
1457 glutamate uptake in organotypic brain slice cultures: Neuroprotection by NF κ B
1458 inhibition. *Brain Res.* 1034, 11–24. <https://doi.org/10.1016/j.brainres.2004.11.014>
- 1459